Asymmetric Syntheses of (–)-ADMJ and (+)-ADANJ: 2-Deoxy-2-amino Analogues of (–)-1-Deoxymannojirimycin and (+)-1-Deoxyallonojirimycin

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Supporting Information



ABSTRACT: The asymmetric syntheses of (–)-ADMJ and (+)-ADANJ, the 2-deoxy-2-amino analogues of (–)-1-deoxymannojirimycin and (+)-1-deoxyallonojirimycin, are described herein. Methodology for the ring-closing iodoamination of bishomoallylic amines followed by in situ ring-expansion (via intramolecular ring-opening of the corresponding aziridinium intermediates with a tethered carbamate moiety) to give oxazolidin-2-ones was initially optimized on a model system. Subsequent application of this methodology to two enantiopure bishomoallylic amines (which were produced via aminohydroxylation of an α,β -unsaturated ester, partial reduction, and reaction of the corresponding aldehyde with vinylmagnesium bromide) also proceeded with concomitant *N*-debenzylation to afford the corresponding diastereoisomerically pure (>99:1 dr) oxazolidin-2-ones. Subsequent deprotection of these enantiopure templates gave (–)-ADMJ and (+)-ADANJ as single diastereoisomers in 16% and 24% overall yield, respectively.

INTRODUCTION

The isosteric replacement of oxygen with nitrogen in compounds displaying useful biological activity is a useful strategy in the search for potential therapeutic agents. Aminosugars are defined as monosaccharides having one hydroxyl group replaced by an amino group¹ (although glycosylamines, where such replacement occurs at the C(1) position, are excluded from this category), and iminosugars are defined as monosaccharides in which the endocyclic oxygen atom has been replaced by a nitrogen atom.² Aminosugars and iminosugars have been a crucial area of recent research with regard to the isosteric replacement of oxygen with nitrogen as naturally occurring deoxyaminosugars and deoxyiminosugars which exhibit useful biological activity are already known. For example, glucose 1, glucosamine 2 (i.e., 2-deoxy-2-aminoglucose), and nojirimycin 3 (i.e., 5-deoxy-5aminoglucose) are illustrative of the isosteric replacement of oxygen with nitrogen: glucosamine 2 has been shown to bring about symptomatic relief of osteoarthritis,³ and nojirimycin 3 displays antimicrobial activity against several drug-resistant strains of bacteria.⁴ The formal replacement of oxygen with nitrogen in compounds already containing at least one amino group furnishes polyamines with further biological applications. For instance, platinum complexes of 3-deoxy-3-aminoglucosamine 4 have been investigated as potential treatments for cancers which offer reduced side effects compared to treatments using cisplatin and carboplatin.⁵ (+)-ADMDP 7, the synthetic 1-deoxy-1-amino analogue of (+)-DMDP 6, and N(1)-substituted derivatives have been reported to display significantly enhanced selectivity and potency toward the inhibition of glucosidases than the parent

iminosugar **6**.⁶ Furthermore, 2-deoxy-2-acetamidonojirimycin **8** (i.e., the *N*-acetyl derivative of 2-deoxy-2-aminonojirimycin **5**) has been investigated as a potent inhibitor of *N*-acetylglucosaminidases,⁷ and 1,2-dideoxy-2-acetamidonojirimycin **9** has been shown to be one of the most powerful reversible inhibitors of hexosaminidases⁸ reported to date (Figure 1). However, relatively few syntheses of deoxyamino analogues of naturally occurring iminosugars have been documented in the literature, with most being derived from carbohydrate precursors.⁹



Figure 1. Structures of several representative aminosugars and iminosugars and their deoxyamino analogues.

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As part of our ongoing research program concerning the asymmetric syntheses of enantiopure pyrrolidines,¹⁰ piperidines,¹¹ and related natural products,¹² we recently reported the ring-closing iodoamination of enantiopure bishomoallylic amine 11, which proceeds with concomitant N-debenzylation, to give iodomethyl pyrrolidine 12. Bishomoallylic amine 11 was prepared in four steps from enantiopure α -hydroxy- β -amino ester 10, which in turn was prepared from the corresponding $\alpha_{\mu}\beta$ -unsaturated ester using our asymmetric aminohydroxylation protocol.¹³ Treatment of bishomoallylic amine 11 with I₂ and NaHCO₃ in MeCN gave iodomethyl pyrrolidine 12, which was then elaborated to 2,5-dideoxy-2,5-imino-D-glucitol [(+)-DGDP] 13 in 6 steps and 65% overall yield, and the 1-deoxy-1-amino analogue 1,2,5-trideoxy-1-amino-2,5-imino-Dglucitol [(+)-ADGDP] 14, which was isolated as the corresponding dihydrochloride salt 14.2HCl, in 23% yield and 99:1 dr.¹⁴ Alternatively, iodomethyl pyrrolidine 12 was converted into carbonate 16 in quantitative yield following intramolecular ring-opening of aziridinium intermediate 15 at the C(5)position by a carbonate group tethered to the C(4) position. However, under optimized conditions the triol derivative 17 was obtained in 40% overall yield (from 11) following sequential ring-closing iodoamination, desilvlation of 16, and methanolysis of the carbonate functionality. Subsequent hydrogenolysis of 17 then gave (-)-1-deoxymannojirimycin [(-)-DMJ] 18 in 87% yield and >99:1 dr (Scheme 1).

Scheme 1



Herein, we describe the extension of this ring-expansion methodology for introduction of other substituents around the piperidine scaffold, specifically targeting the 2-deoxy-2-amino analogues of (-)-1-deoxymannojirimycin [(-)-DMJ] **18** and (+)-1-deoxyallonojirimycin [(+)-DANJ] **24**. This methodology was initially explored in a model system, where bishomoallylic amine **19** was first subjected to the ring-closing iodoamination protocol in the presence of CO₂ (for the formation of cyclic carbonate **21**); then alternative "X=C=Y" electrophiles (e.g., X, Y = O, NR, S, etc.) were examined for the formation of **23**. The application of this strategy to enantiopure bishomoallylic amine **11** and its epimer then culminated in the total asymmetric syntheses of (-)-ADMJ **25** and (+)-ADANJ **26**, the 2-deoxy-2-amino analogues of (-)-DMJ **18** and (+)-DANJ **24**, respectively (Figure 2).



Figure 2. Synthetic strategy toward (–)-ADMJ 25 and (+)-ADANJ 26, the 2-deoxy-2-amino analogues of (–)-DMJ 18 and (+)-DANJ 24.

RESULTS AND DISCUSSION

The model bishomoallylic amine substrate 19 was prepared via monobenzylation of amino alcohol 27 upon treatment with benzaldehyde and CH(OMe)₃ followed by NaBH₄, which gave N-benzyl-substituted amino alcohol 28 in 90% yield.¹⁶ Subsequent reaction of 28 with Boc₂O in a mixture of CH₂Cl₂ and 1.0 M aq NaOH (5:1) at 0 °C delivered N-Boc-N-benzylprotected amino alcohol 29 in 95% vield.¹⁷ Oxidation of the primary hydroxyl moiety within 29 under Swern conditions followed by direct treatment of the crude reaction mixture of aldehyde 30 with vinylmagnesium bromide gave N-Boc-protected bishomoallylic amine 31 in 46% yield over two steps (from 29). Subjection of 31 to 1.25 M HCl in MeOH at 40 °C effected N-Boc deprotection, and N-benzyl-substituted bishomoallylic amine 19 was then isolated in 74% yield. Repetition of this procedure while omitting the purification of 31 reproducibly led to the production of 19 in 66% isolated yield over three steps (from 29) on multigram scales (Scheme 2).

Ring-closing iodoamination/ring-expansion of **19** using the conditions that were previously¹⁵ optimized for the conversion of bishomoallylic amine **11** into the corresponding carbonate **16** resulted in the formation of cyclic carbonate **32**. Although **32** was observed as the major compound in the ¹H NMR spectrum of the crude reaction mixture, it was isolated in only 36% yield after flash column chromatography.¹⁸ Methanolysis of an analytically pure sample of **32** upon treatment with K_2CO_3 and MeOH gave diol **33** in quantitative yield. Analysis of the ¹H NMR ³J coupling constants within **33** proved difficult due to the peaks in the ¹H NMR spectrum of **33** (recorded at rt





in CDCl₃) being broad.¹⁹ Thus, the relative configurations within diol 33 and the carbonate precursor 32 were established via chemical correlation. It was anticipated that tetrahydropyridine 34 would be the ideal common precursor for the syntheses of authentic samples of both *cis*-diol 33^{20} and *trans*-diol 36^{21} (and the corresponding carbonates cis-32 and trans-37). The preparation of 34 was achieved via the reduction of *N*-benzylpyridinium bromide following a literature procedure:²⁴ treatment of pyridine with BnCl at 140 °C followed by reduction of N-benzylpyridinium bromide with NaBH₄ gave tetrahydropyridine 34^{22} in 81% yield. Stereospecific *cis*-dihydroxylation²³ of 34 upon treatment with OsO₄ and NMO (i.e., under Upjohn conditions²⁴) in a mixture of THF/H₂O (5:1) gave *cis*-diol 33, which was isolated in 67% yield after purification via flash column chromatography. Alternatively, epoxidation of the corresponding ammonium salt of 34 (to prevent N-oxidation) using HBF₄ and *m*-CPBA in CH_2Cl_2 produced epoxide 35,² which underwent reaction with H_2SO_4 in dioxane/ H_2O (40:1) to give trans-diol 36 in 56% overall yield (from 34). Authentic samples of cis-carbonate 32 and trans-carbonate 37 were then synthesized from cis-diol 33 and trans-diol 36, respectively: treatment of cis-diol 33 with CDI and DMAP in THF promoted full conversion to cis-carbonate 32, which was observed as the sole product of the reaction in the ¹H NMR spectrum of the crude reaction mixture and was subsequently isolated in 36% yield after purification via flash column chromatography. The preparation of trans-carbonate 37 using similar conditions required 4 days to reach completion, and 37 was characterized as the sole product of the reaction, as determined by ¹H NMR spectroscopic analysis of the crude reaction mixture. Attempted purification of trans-37 by flash column chromatography led to its degradation; therefore, ¹H and ¹³C NMR spectroscopic analyses were recorded using the crude reaction mixture. Comparison of the ¹H NMR spectra of these authentic samples of cis-carbonate 32 and trans-carbonate 37 (and cis-diol 33 and trans-diol 36) with the ¹H NMR spectrum of the ring-closing iodoamination/ring-expansion product 32 derived from bishomoallylic amine 19 established the relative configurations within both 32 and 33 and also confirmed the absence of the alternative diastereoisomeric products 36 and 37 from the reaction of 19 under the ring-closing iodoamination conditions (Scheme 3).



Having established that the conditions developed for the trapping of CO₂ could be applied in the one-pot ring-closing iodoamination/ring-expansion of the model bishomoallylic substrate 19, a study into the mechanism of reaction was next initiated. Iodoamination of 19 upon treatment with 3.0 equiv of I₂ and 3.0 equiv of NaHCO₃ in MeCN resulted in the formation of a 70:30 mixture of iodopiperidines 38 and 39, respectively. Although the crude reaction mixture was obtained in quantitative mass return, the purification of 38 and 39 by flash column chromatography proved difficult and only 38 could be isolated as a pure sample in 24% yield, and an enriched sample of 39 (70:30 dr) was collected in 12% yield (Scheme 4). The relative configuration of 38 was established unambiguously by single-crystal X-ray diffraction analysis.²⁵ ¹H and ¹³C NMR spectroscopic analyses, including a ¹H-¹³C HMBC study, supported the assigned connectivity within 39. The chemical shifts of the carbons directly bonded to iodine in the ¹³C NMR spectra of both 38 (δ_c = 39.9 ppm) and 39 (δ_c = 38.4 ppm) were diagnostic of the iodine being supported by a CH carbon (i.e., a piperidine scaffold) as opposed to a CH₂ carbon (i.e., a pyrrolidine scaffold).²⁶ The ¹H NMR ³J coupling constant analyses of these samples of 38 and 39 were also consistent with their assigned relative configurations. In contrast to the sharp peaks displayed in the ¹H NMR spectrum of 39 (recorded at rt in CDCl₃), the peaks observed in the ¹H NMR spectrum of 38 (recorded at rt in CDCl₃) were extremely broad²⁷ and only a limited number of correlations could be distinguished in the ¹H-¹H COSY, ¹H-¹³C HSQC, and ¹H-¹³C HMBC NMR spectra. The ¹H and ¹³C NMR spectra of **38** were therefore recorded at 363 K in PhMe- d_8 . In this case the peaks in the ¹H and ¹³C NMR spectra were resolved, and characteristic correlations were also observed in the corresponding 2D NMR spectra; these data were all consistent with the structure of 38. The formation of iodopiperidines 38 and 39

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in the iodoamination of 19 is in direct contrast with the formation of iodomethyl pyrrolidine 12 when bishomoallylic amine 11 was subjected to the same conditions.¹⁵ This reaction outcome could be rationalized by either 5-exo cyclization of the corresponding iodonium species followed by rearrangement (presumably via the intermediacy of the corresponding aziridinium species) or 6-endo cyclization. Repetition of the iodoamination of 19 followed by direct treatment of the 70:30 crude reaction mixture of 38 and 39 with NaHCO₃ in a mixture of dioxane/H₂O (3:1) afforded a 70:30 mixture of carbonate 32 and iodopiperidine 39. Subsequent purification by flash column chromatography led to the isolation of 32 in 28% yield and 39 in 5% yield as single diastereoisomers (>99:1 dr) in each case (Scheme 4). The isolation of iodopiperidine 39 following this stepwise process was in contrast with the sole isolation of carbonate 32 when performing the corresponding one-pot transformation (vide supra) and provided some insight into the reaction mechanism: in this case, carbonate 32 seemed to derive from 38, while 39 is apparently inert under the conditions for carbonate formation.

With diastereoisomerically pure (>99:1 dr) samples of iodopiperidines cis-38 and trans-39 in hand, it was now possible to separately investigate the trapping of CO₂ from NaHCO₃ by each diastereoisomer. Treatment of 38 with AgBF₄ in CD₂Cl₂ delivered the corresponding aziridinium 40, which could not be isolated due to degradation. Aziridinium 40 was subsequently subjected to the trapping conditions (i.e., NaHCO₃ in a 3:1 mixture of dioxane/H₂O was added to the NMR sample) to give carbonate 32 in quantitative yield as a single diastereoisomer (>99:1 dr). Direct treatment of 38 with NaHCO₃ in a mixture of dioxane/ H_2O (3:1) promoted the formation of carbonate 32, which was obtained in quantitative yield and >99:1 dr, thereby supporting the intermediacy of aziridinium 40 in the conversion of iodopiperidine 38 to carbonate 32. In contrast, 39 was converted into the corresponding aziridinium 41 (upon treatment of **39** with AgBF₄ in CD_2Cl_2) and subjected to the same trapping conditions, which gave a 77:15:8 mixture of pyrrolidine 42, piperidine 36, and carbonate 32, respectively. Presumably, intermolecular ring-opening of 41 by H₂O gave pyrrolidine 42 [i.e., ring-opening at C(6)] and piperidine 36 [i.e., ring-opening at C(5) with inversion of configuration]. The formation of carbonate 32 in this case was rationalized by the trapping of CO₂ by *trans*-iodopiperidine 39 and subsequent S_N2-type substitution of the C(5)-iodide within 43 (Scheme 5). Direct treatment of 39

Scheme 5



with NaHCO₃ in a mixture of dioxane/ $H_2O(3:1)$ resulted in the formation of a complex mixture of products.

Having demonstrated that aziridinium 40 successfully underwent ring-expansion via the trapping of CO_2 to give carbonate 32, our attention next turned to examine the trapping of sulfurand nitrogen-containing "X=C=Y" electrophiles. It was envisaged that the trapping of either isothiocyanates (RNCS) or isocyanates (RNCO) by aziridinium 40 would result in the subsequent intramolecular ring-opening at the C(5) position by the more nucleophilic sulfur or nitrogen atom, respectively, to give the corresponding oxathiolan-2-one (after hydrolysis of the oxathiolan-2-imine intermediate) or oxazolidin-2-one. Prior to investigating the trapping of isothiocyanates and isocyanates, it was anticipated that the synthesis of authentic samples of oxathiolan-2-one 46 and oxazolidin-2-one 47 could be achieved by the intermolecular ring-opening of aziridinium 40 at the C(5) position by either the thiocyanate anion (NCS⁻) or the cyanate anion (OCN⁻), respectively, followed by cyclization. Thus, 38 was treated with AgSCN, which resulted in the formation of an 85:15 regioisomeric mixture of bicyclic derivatives 44 and 45, from which 44 was isolated in 43% yield and >99:1 dr and 45 was isolated in 4% yield and >99:1 dr (Scheme 6). The atomic connectivities within 44 and 45 were assigned by ¹H and ¹³C NMR spectroscopic analyses, including ¹H-¹³C

Scheme 6



HMBC studies. In addition, characteristic C=N absorbances at 1639 cm^{-1} in the IR spectra of both 44 and 45, along with diagnostic chemical shifts for the SCN carbons ($\delta_{\rm C} = \sim 169$ ppm) in the ¹³C NMR spectra of both 44 and 45, confirmed the presence of a carbonimidothioate moiety. Hydrolysis of 44 upon treatment with 1.0 M aq HCl delivered oxathiolan-2-one 46 in 39% yield and >99:1 dr. The IR spectrum of 46 displayed a diagnostic C=O absorbance at 1732 cm⁻¹, which along with the diagnostic chemical shift at $\delta_{\rm C}$ = 172.5 ppm for the SCO carbon observed in the ¹³C NMR spectrum of 46 fully supported the assigned structure of 46. Analogous reaction of 38 with AgOCN afforded an 85:15 regioisomeric mixture of bicyclic derivatives 47 and 48, from which 47 was isolated in 53% yield and >99:1 dr and 48 was isolated in 7% yield and >99:1 dr (Scheme 6). The atomic connectivities within 47 and 48 were again assigned following ¹H and ¹³C NMR spectroscopic analyses, including ¹H-¹³C HMBC studies. In addition, the presence of diagnostic C=O absorbances at 1749 cm^{-1} in the IR spectrum of 47 and at 1756 cm⁻¹ in the IR spectrum of 48 were entirely consistent with the presence of a carbamate functionality. In both cases, ring-opening of aziridinium 40 by thiocyanate or cyanate anions at the C(5) position (with inversion of configuration) was responsible for the formation of the major regioisomers 44 and 47, while ring-opening of 40 at the C(6) position was responsible for the formation of the minor regioisomers 45 and 48.

The trapping of either isothiocyanates (RNCS) or isocyanates (RNCO) was investigated next. Treatment of **38** with AgBF₄ in CH₂Cl₂ (to convert **38** to the corresponding aziridinium **40**) followed by the addition of PhNCS gave a complex mixture of products. Alternatively, reaction of **38** with either PhNCS or BzNCS followed by AgBF₄ also resulted in the formation of complex mixtures of products, failing to give oxathiolan-2-imines **49** or **50** (or the corresponding oxathiolan-2-ones) in all attempts. In light of these results, the trapping of isothiocyanates was abandoned and attention was next turned to the trapping of isocyanates. While treatment of **38** with PhNCO followed by

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the addition of AgBF₄ did not provide oxazolidin-2-one 54, submission of 38 to the same reaction conditions with BzNCO resulted in the formation of oxazolidin-2-one 55, which was observed as the major product in the ¹H NMR spectrum of the crude reaction mixture. In this case, purification by flash column chromatography gave 55 in 23% yield as a single diastereoisomer (>99:1 dr). Similarly, treatment of 38 with p-toluenesulfonyl isocyanate (TsNCO) followed by the addition of AgBF₄ produced oxazolidin-2-one 56, which was observed as the sole product upon inspection of the ¹H NMR spectrum of the crude reaction mixture and obtained in 62% yield and >99:1 dr after purification by flash column chromatography.²⁸ The relative configuration of 56 was unambiguously established by single-crystal X-ray diffraction analysis.²⁵ The relative configuration of 55 was assigned by analogy [on the basis that the ring-opening of aziridinium 52 proceeds with inversion of configuration at the C(5) position], and this assignment was supported via ¹H NMR ³J coupling constant correlation with both 47 and 56. Subjecting 39 to TsNCO in CH₂Cl₂ followed by the addition of AgBF₄ led to the isolation of the epimeric oxazolidin-2-one 57 in 30% yield and >99:1 dr (Scheme 7).²





It was interesting to note that the formation of **57** from the formal trapping of TsNCO by **39** was in direct contrast with the unsuccessful formation of *trans*-carbonate **37** from **39**. While the conversion of **38** to **49** or **50** was unsuccessful, the trapping of TsNCO led to the isolation of oxazolidin-2-one **56** in good yield as a single regioisomer, and it was envisaged that this methodology could be employed in the syntheses of (-)-ADMJ **25** and (+)-ADANJ **26**, the 2-deoxy-2-amino analogues of (-)-DMJ **18** and (+)-DANJ **24**.

Treatment of a freshly prepared sample of iodomethylpyrrolidine **12** with TsNCO in CH_2Cl_2 for 5 min followed by the addition of AgBF₄ produced a 55:40:5 mixture of dioxolan-2-imine



59 and carbamates 60 and 61, respectively, in quantitative mass return (Scheme 8). This outcome is consistent with the formation of aziridinium ion 58 followed by tethered ringopening at the C(5) position through either the carbamate oxygen atom (to give dioxolan-2-imine 59) or the carbamate nitrogen atom (to give cyclic carbamate 60). Carbamate 61 is presumably derived from a trace amount of the corresponding N(1)- α -methylbenzyl-substituted pyrrolidine 62 present in the sample of 12.³⁰ The IR spectrum of the crude reaction mixture displayed a diagnostic C=N absorbance at 1640 cm⁻¹, supporting the assigned identity of 59, and upon attempted purification of the crude reaction mixture by flash column chromatography carbonate 16 (which presumably resulted from hydrolysis of 59) was recovered in 48% yield. A 90:10 mixture of carbamates 60 and 61 was also isolated, and the IR spectrum of this mixture displayed a diagnostic C=O absorbance at 1788 cm⁻¹. The relative configuration of **60** (and **61**) was initially assigned by analogy to the stereochemical outcome observed upon reaction of the model substrate 39 under these conditions and was later confirmed via ¹H NMR ³J coupling constant analyses of several derivatives.

Further reaction optimization revealed that treatment of bishomoallylic amine 11 with I₂ and NaHCO₃ in MeCN followed by the addition of 4.5 equiv of TsNCO to the reaction mixture after 16 h (i.e., omitting the addition of AgBF₄) gave a 76:19:5 mixture of **60**, **63**, and **61**, respectively, in addition to N-(α -methylbenzyl)acetamide (which resulted from loss of the N- α -methylbenzyl group in a Ritter-type process).¹⁵ Purification by flash column chromatography resulted in the

isolation of a 76:19:5 mixture of **60**, **63**, and **61**, respectively. Repetition of the reaction followed by direct methanolysis of the crude reaction mixture upon treatment with K_2CO_3 and MeOH gave a 76:19:5 mixture of **64**, **65**, and **66**, respectively. Purification of the crude reaction mixture by flash column chromatography gave **64** in 25% yield (from **11**), **65** in 7% yield (from **11**), and **66** in 2% yield (from **11**) as single diastereoisomers (>99:1 dr) in each case (Scheme 9). The relative





configurations of piperidines **64–66** (and therefore also those of the synthetic precursors **60**, **61**, and **63**) were established by ¹H NMR ³J coupling constant analyses, and the absolute configurations of these compounds were assigned from the known absolute configurations at the C(2), C(3), C(4), and C(α) stereogenic centers within the precursor bishomoallylic amine **11**.

One-pot ring-closing iodoamination/ring-expansion of the epimeric bishomoallylic amine 67^{15} upon treatment with I₂ and NaHCO₃ followed by the addition of TsNCO after 16 h resulted in the formation of carbamate 68 and *N*-(α -methylbenzyl)-acetamide. Purification of the crude reaction mixture by flash column chromatography gave 68 in 48% yield and >99:1 dr. Subsequent treatment of 68 with K₂CO₃ and MeOH led to the isolation of piperidine 69 in 83% yield and >99:1 dr (Scheme 10). The relative configuration of 69 was unambiguously established by single-crystal X-ray diffraction analysis.²⁵

Scheme 10



Furthermore, the determination of a Flack x parameter³¹ of -0.004(9) for the crystal structure of **69** allowed the assigned absolute (2*R*,3*R*,4*S*,5*S*)-configuration of **69** to be confirmed. This analysis also unambiguously established the absolute configuration of the carbamate precursor **68**.

With piperidines 64 and 69 in hand, it was now possible to commence investigations into the deprotection of the N-tosyl moiety. Following a literature procedure,³² treatment of **69** with Na and naphthalene in THF at -10 °C for 3 h proceeded with concomitant O-benzyl deprotection to give a 40:60 mixture of 72 and 73, respectively. Purification of the crude reaction mixture by flash column chromatography gave 72 in 40% yield and 73 in 60% yield in >99:1 dr in both cases. Reaction optimization revealed that treatment of 69 with Na and naphthalene in THF at rt for 16 h gave a 93:7 mixture of 72 and 73, respectively. Purification of this mixture via flash column chromatography led to the isolation of 72 in 61% yield and >99:1 dr and 73 in 6% yield and >99:1 dr. Submission of 64 to these optimized reaction conditions resulted in the formation of a 95:5 mixture of 70 and 71, from which 70 was isolated in 63% yield and >99:1 dr and 71 was isolated in 4% yield and 99:1 dr (Scheme 11).

Scheme 11



Treatment of **70** with 6.0 M aq HCl in MeOH at 40 °C effected *O*-TIPS deprotection, and subsequent hydrogenolysis upon treatment with Pearlman's catalyst under H₂ (5 atm) followed by purification of this sample by ion exchange chromatography on Dowex 50WX8 (H⁺ form) resin afforded (–)-ADMJ **25** in quantitative yield and >99:1 dr. Identical treatment of **72** gave (+)-ADANJ **26** in quantitative yield and >99:1 dr (Scheme 12). While there was no precedent in the literature for the synthesis or identification of (+)-ADANJ **26**, the specific rotation and ¹H and ¹³C NMR spectra of this sample of (–)-ADMJ **25** { $[\alpha]_D^{20}$ –11.5 (*c* 0.3 in H₂O)} were in very good agreement with the data previously reported for a synthetic sample of (–)-ADMJ **25** by Le Merrer et al.^{9b} { $[\alpha]_D^{-14}$ (*c* 0.4 in H₂O)}.

CONCLUSION

In conclusion, the methodology for the ring-closing iodoamination of bishomoallylic amines followed by in situ ring-expansion (via intramolecular ring-opening of the corresponding aziridinium intermediates with a tethered carbamate moiety) to give oxazolidin-2-ones was initially optimized on a model system. Subsequent application of this methodology to two enantiopure bishomoallylic amines (which were produced via aminohydroxylation of an $\alpha_i\beta$ -unsaturated ester, partial reduction,





and reaction of the resultant aldehyde with vinylmagnesium bromide) also proceeded with concomitant *N*-debenzylation to afford the corresponding diastereoisomerically pure (>99:1 dr) oxazolidin-2-ones. Subsequent deprotection of these enantiopure templates gave (–)-ADMJ and (+)-ADANJ, the 2-deoxy-2-amino analogues of (–)-1-deoxymannojirimycin and (+)-1-deoxyallonojirimycin, as single diastereoisomers (>99:1 dr) in 16% and 24% overall yield, respectively.

EXPERIMENTAL SECTION

General Experimental Details. All reactions involving organometallic or other moisture-sensitive reagents were carried out under a nitrogen atmosphere using standard vacuum line techniques and glassware that was flame dried and cooled under nitrogen before use. Solvents were dried according to the procedure outlined by Grubbs and co-workers.³³ BuLi was purchased as a solution in hexanes and titrated against diphenylacetic acid before use. All other reagents were used as supplied without prior purification. Organic layers were dried over Na₂SO₄. Thin layer chromatography was performed on aluminum plates coated with 60 F_{254} silica. Plates were visualized using UV light (254 nm), 1% aq KMnO₄, or Dragendorff's reagent. Flash column chromatography was performed on Kieselgel 60 silica. Melting points are uncorrected. Specific rotations are reported in 10^{-1} deg cm² g⁻¹ and concentrations in g/100 mL. IR spectra were recorded using an ATR module. Selected characteristic peaks are reported in cm⁻¹. NMR spectra were recorded in the deuterated solvent stated. Spectra were recorded at rt unless otherwise stated. The field was locked by external referencing to the relevant deuteron resonance. ${}^{1}H-{}^{1}H$ COSY, ¹H-¹³C HMQC, and ¹H-¹³C HMBC analyses were used to establish atom connectivity. Accurate mass measurements were run on a TOF spectrometer internally calibrated with polyalanine. X-ray Crystal Structure Determination.²⁵ Data were

X-ray Crystal Structure Determination.²⁵ Data were collected using either graphite-monochromated Mo/K α radiation (for 39) or graphite-monochromated Cu/K α radiation (for 56 and 69) using standard procedures at 150 K. The structure was solved by direct methods (SIR92); all non-hydrogen atoms were refined with anisotropic thermal parameters. Hydrogen atoms were added at idealized positions. The structure was refined using CRYSTALS.³⁴

3-(N-Benzylamino)propan-1-ol 28. 3-Aminopropan-1-ol 27 (5.00 g, 66.6 mmol) was added to a stirred solution of PhCHO (6.77 mL, 66.6 mmol) and CH(OMe)₃ (10.9 mL, 99.8 mmol), and the resultant mixture was stirred at rt for 5 h. The reaction mixture was then cooled to 0 °C, and NaBH₄ (2.52 g, 66.6 mmol) was added portionwise. The reaction mixture was then concentrated in vacuo, and the residue was partitioned between H₂O (200 mL) and EtOAc (200 mL). The aqueous layer was extracted with EtOAc (2 × 50 mL), and the combined organic extracts were then dried and concentrated in vacuo to give 28 as a colorless oil (9.95 g, 90%);¹⁶ $\delta_{\rm H}$ (400 MHz,

CDCl₃) 1.76 (2H, app quintet, J 5.5, C(2) H_2), 2.93 (2H, t, J 5.5, C(3) H_2), 3.16 (2H, br s, OH, NH), 3.82 (2H, s, NC H_2 Ph), 3.84 (2H, t, J 5.5, C(1) H_2), 7.26–7.38 (5H, m, Ph).

3-[*N*-**Benzyl-***N***-(***tert***-butoxycarbonyl)amino]propan-1-ol 29**. (Boc)₂O (6.60 g, 30.3 mmol) was added to a stirred solution of **28** (5.00 g, 30.3 mmol) in CH₂Cl₂/1.0 M aq NaOH (5:1, 90 mL) at 0 °C. The resultant mixture was allowed to warm to rt and stirred at rt for 16 h. The reaction mixture was washed with H₂O (100 mL), dried, and concentrated in vacuo to give **29** as a colorless oil (7.66 g, 95%);^{17,35} $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.49 (9H, s, CM ϵ_3), 1.65 (2H, app br s, C(2)H₂), 2.75 (1H, br s, OH), 3.40 (2H, app br s, C(3)H₂), 3.57–3.61 (2H, m, C(1)H₂), 4.41 (2H, br s, NCH₂Ph), 7.23–7.38 (5H, m, *Ph*).

5-[*N*-Benzyl-*N*-(*tert*-butoxycarbonyl)amino]pent-1-en-3-ol **31.** *Step 1.* DMSO (0.24 mL, 3.3 mmol) was added dropwise to a stirred solution of $(COCl)_2$ (0.12 mL, 1.4 mmol) in CH_2Cl_2 (15 mL) at -78 °C. After 20 min, a solution of **29** (200 mg, 0.75 mmol) in CH_2Cl_2 (15 mL) at -78 °C was added dropwise via cannula. After a further 20 min, Et₃N (0.63 mL, 4.5 mmol) was added and the resultant mixture was stirred at -78 °C for 30 min before being allowed to warm to rt over 30 min. The reaction mixture was then concentrated in vacuo, and the residue was partitioned between H₂O (30 mL) and Et₂O (30 mL). The aqueous layer was extracted with Et₂O (2 × 15 mL), and the combined organic extracts were then dried and concentrated in vacuo to give **30** as a yellow oil (204 mg); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.48 (9H, s, CMe₃), 2.60-2.72 (2H, m, C(2)H₂), 3.39-3.56 (2H, m, C(3)H₂), 4.40-4.47 (2H, m, NCH₂Ph), 7.25-7.37 (5H, m, Ph), 9.75 (1H, s, C(1)H).

Step 2. Vinylmagnesium bromide (1.0 M in THF, 2.25 mL, 2.25 mmol) was added dropwise to a stirred solution of the residue of 30 (204 mg) in THF (5 mL) at 0 °C. The reaction mixture was allowed to warm to rt and stirred at rt for 16 h. The reaction mixture was then cooled to 0 °C, and H2O (1 mL) was added dropwise. The resultant mixture was concentrated in vacuo, and the residue was partitioned between H₂O (10 mL) and EtOAc (10 mL). The aqueous layer was extracted with EtOAc $(2 \times 5 \text{ mL})$, and the combined organic extracts were washed with brine (10 mL), dried, and concentrated in vacuo. Purification via flash column chromatography (eluent 30-40 °C petrol/Et₂O, 2:1) gave 31 as a yellow oil (100 mg, 46% from 29); ν_{max} (ATR) 3428 (O-H), 2976, 2932 (C-H), 1692, 1670 (C=O), 1477, 1454, 1417 (C=C); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.44–1.60 (10H, m, CMe_{3} , C(4) H_{A}), 1.68–1.76 (1H, m, C(4) H_{B}), 3.06–3.10 (1H, m, $C(5)H_A$, 3.72–3.76 (1H, m, $C(5)H_B$), 4.01 (1H, br s, OH), 4.06–4.10 (1H, m, C(3)H), 4.26–4.31 (1H, m, NCH_AH_BPh), 4.53 (1H, d, J 15.6, NCH_AH_BPh), 5.08 (1H, d, J 10.4, C(1)H_A), 5.23 (1H, d, J 17.2, $C(1)H_B$, 5.85 (1H, ddd, J 17.2, 10.4, 5.2, C(2)H), 7.21–7.36 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 28.4 (CMe₃), 35.2 (C(4)), 42.7 (C(5)), 50.6 (NCH₂Ph), 68.7 (C(3)), 80.5 (CMe₃), 113.8 (C(1)), 127.3 (p-Ph), 128.5, 128.6 (o,m-Ph), 138.1 (i-Ph), 140.4 (C(2)), 156.8 (NCO); m/z (ESI⁺) 292 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{17}H_{26}NO_3^+$ ([M + H]⁺) requires 292.1907; found 292.1908.

5-(N-Benzylamino)pent-1-en-3-ol 19. Method A (from 31). A solution of 31 (50 mg, 0.17 mmol) in HCl (1.25 M in MeOH, 2 mL) was heated at 40 °C for 16 h before being allowed to cool to rt and concentrated in vacuo. The residue was then partitioned between 2.0 M aq KOH (10 mL) and CHCl₃/ⁱPrOH (3:1, 10 mL). The aqueous layer was extracted with $CHCl_3$ /ⁱPrOH (3:1, 2 × 5 mL), and the combined organic extracts were then dried over Na2SO4 and concentrated in vacuo to give 19 as a yellow oil (24 mg, 74%); $\nu_{\rm max}$ (ATR) 3298 (N-H, O-H), 2924, 2850 (C-H), 1495, 1454 (C=C); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.60–1.70 (1H, m, C(4)H_A), 1.76–1.84 (1H, m, $C(4)H_B$), 2.86 (1H, ddd, J 12.3, 9.1, 3.5, $C(5)H_A$), 3.01 (1H, ddd, J 12.3, 6.4, 3.5, C(5)H_B), 3.65 (1H, br s, OH), 3.80 (1H, d, J 13.0, NCH_AH_BPh), 3.84 (1H, d, J 13.0, NCH_AH_BPh), 4.32-4.39 (1H, m, C(3)H), 5.08 (1H, app dt, J 10.5, 1.5, C(1)H_A), 5.27 (1H, app dt, J 17.0, 1.5, $C(1)H_B$, 5.85 (1H, ddd, J 17.0, 10.5, 5.2, C(2)H), 7.25-7.36 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 34.4 (C(4)), 47.1 (C(5)), 53.4 (NCH₂Ph), 73.5 (C(3)), 113.9 (C(1)), 127.5 (*p*-Ph), 128.4, 128.6 (o,m-Ph), 138.2 (i-Ph), 140.7 (C(2)); m/z (ESI⁺) 192 $([M + H]^+, C(2))$ 100%); HRMS (ESI⁺) C₁₂H₁₈NO⁺ ([M + H]⁺) requires 192.1383; found 192.1387.

Method B (from 29). Step 1: DMSO (5.90 mL, 83.0 mmol) was added dropwise to a stirred solution of $(COCl)_2$ (2.92 mL, 34.0 mmol) in CH₂Cl₂ (400 mL) at -78 °C. After 20 min, a solution of 29 (5.00 g, 18.9 mmol) in CH₂Cl₂ (400 mL) at -78 °C was added dropwise via cannula. After a further 20 min, Et₃N (15.8 mL, 114 mmol) was added and the resultant mixture was stirred at -78 °C for 30 min before being allowed to warm to rt over 30 min. The reaction mixture was then concentrated in vacuo, and the residue was partitioned between H₂O (500 mL) and Et₂O (500 mL). The aqueous layer was extracted with Et₂O (2 × 100 mL), and the combined organic extracts were then dried and concentrated in vacuo to give 30 as a yellow oil (5.26 g).

Method B (from 29). Step 2: Vinylmagnesium bromide (1.0 M in THF, 56.7 mL, 56.7 mmol) was added dropwise to a stirred solution of the residue of 30 (5.26 g) in THF (400 mL) at 0 °C. The reaction mixture was allowed to warm to rt and stirred at rt for 16 h. The reaction mixture was then cooled to 0 °C, and H₂O (20 mL) was added dropwise. The resultant mixture was concentrated in vacuo, and the residue was partitioned between H₂O (400 mL) and EtOAc (400 mL). The aqueous layer was extracted with EtOAc (2 × 50 mL), and the combined organic extracts were washed with brine (400 mL), dried, and concentrated in vacuo to give 31 as a yellow oil (5.59 g).

Method B (from **29**). Step 3: The residue of **31** (5.59 g) was dissolved in HCl (1.25 M in MeOH, 50 mL), and the resultant mixture was heated at 40 °C for 16 h before being allowed to cool to rt and concentrated in vacuo. The residue was partitioned between 2.0 M aq KOH (50 mL) and CHCl₃/ⁱPrOH (3:1, 50 mL). The aqueous layer was extracted with CHCl₃/ⁱPrOH (3:1, 2 × 20 mL), and the combined organic extracts were then dried over Na₂SO₄ and concentrated in vacuo. Purification via flash column chromatography (eluent CHCl₃/ MeOH, 48:1) gave **19** as a yellow oil (2.40 g, 66% from **29**).

(RS,SR)-N(1)-Benzyl-3,4-dihydroxy-3,4-O-carbonylpiperidine 32. Method A (from 19). I_2 (1.20 g, 4.71 mmol) and NaHCO₃ (396 mg, 4.71 mmol) were added to a stirred solution of 19 (300 mg, 1.57 mmol) in dioxane/H2O (3:1, 4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was then diluted with Et₂O (15 mL), washed with satd aq Na₂S₂O₃ (15 mL), dried, and concentrated in vacuo. Purification via flash column chromatography (eluent 30-40 °C petrol/EtOAc, 2:1) gave 32 as a yellow oil (130 mg, 36%, >99:1 dr); ν_{max} (ATR) 2925 (C–H), 1799 (C=O); δ_{H} (400 MHz, CDCl₃) 1.98-2.07 (1H, m, C(5)H_A), 2.17 (1H, app dq, J 15.1, 4.2, $C(5)H_B$, 2.42–2.56 (3H, m, $C(2)H_A$, $C(6)H_2$), 2.94 (1H, ddd, J 12.5, 5.4, 1.4, C(2)H_B), 3.56 (1H, d, J 13.1, NCH_AH_BPh), 3.58 (1H, d, J 13.1, NCH_AH_BPh), 4.69 (1H, app q, J 6.4, C(3)H), 4.73-4.78 (1H, m, C(4)H), 7.25–7.38 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 26.4 (C(5)), 47.3 (C(6)), 53.3 (C(2)), 62.2 (NCH₂Ph), 73.7 (C(3)), 74.1 (C(4)), 127.4 (*p*-*Ph*), 128.4, 128.8 (*o*,*m*-*Ph*), 137.2 (*i*-*Ph*), 155.0 (CO); m/z (ESI⁺) 234 ([M + H]⁺, 100%); HRMS (ESI⁺) C₁₃H₁₅NNaO₃⁺ $([M + Na]^{+})$ requires 256.0944; found 256.0950.

Method B (from 33). CDI (59 mg, 0.36 mmol) and DMAP (6 mg, 0.05 mmol) were added to a stirred solution of 33 (50 mg, 0.24 mmol, >99:1 dr) in THF (2 mL), and the resultant mixture was allowed to stir at rt for 24 h. Saturated aq NH₄Cl (0.5 mL) was added, and the reaction mixture was extracted with EtOAc (2×5 mL). The combined organic extracts were washed with brine (10 mL), dried, and concentrated in vacuo. Purification via flash column chromatography (eluent 30–40 °C petrol/EtOAc, 2:1) gave 32 as a yellow oil (20 mg, 36%, >99:1 dr).

Method C (from **38**). AgBF₄ (18 mg, 95 μ mol) was added to **38** (25 mg, 78 μ mol, >99:1 dr) in CD₂Cl₂, and the reaction mixture was shaken at rt for 5 min. The reaction mixture was then poured into a stirred solution of NaHCO₃ (20 mg, 0.23 mmol) in dioxane/H₂O (3:1, 4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was then diluted with Et₂O (10 mL), and the organic layer was washed with satd aq NaHCO₃ (10 mL), dried, and concentrated in vacuo to give **32** as a yellow oil (20 mg, quant, >99:1 dr).

(*RS,SR*)-*N*(1)-Benzyl-3,4-dihydroxypiperidine 33. *Method A* (*from 32*). K_2CO_3 (116 mg, 0.84 mmol) was added to a stirred solution of 32 (65 mg, 0.28 mmol, >99:1 dr) in MeOH (4 mL) at rt, and the resultant mixture was allowed to stir at rt for 16 h before being

concentrated in vacuo. The residue was then partitioned between H₂O (5 mL) and CHCl₃/ⁱPrOH (3:1, 5 mL), and the aqueous layer was extracted with CHCl₃/ⁱPrOH (3:1, 2 × 5 mL). The combined organic extracts were then dried and concentrated in vacuo to give **33** as a yellow oil (60 mg, quant, >99:1 dr);²⁰ ν_{max} (ATR) 3377 (O–H), 2812, 2926 (C–H); $\delta_{\rm H}$ (400 MHz, CD₂Cl₂) 1.63–1.72 (1H, m, C(5)H_A), 1.73–1.78 (1H, m, C(5)H_B), 2.04–2.11 (1H, m, C(6)H_A), 2.26 (1H, app d, J 10.6, C(2)H_A), 2.65–2.72 (1H, m, C(6)H_B), 2.80 (1H, app br s, C(2)H_B), 3.50 (1H, d, J 13.2, NCH_AH_BPh), 3.53 (2H, m, NCH_AH_BPh, C(4)H), 3.69–3.71 (1H, m, C(3)H), 7.23–7.33 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CD₂Cl₂) 30.5 (C(5)), 50.9 (C(6)), 57.5 (C(2)), 62.6 (NCH₂Ph), 69.3 (C(3)), 69.8 (C(4)), 127.7 (*p*-Ph), 128.8, 129.5 (*o*,*m*-Ph), 138.8 (*i*-Ph); *m*/z (ESI⁺) 208 ([M + H]⁺, 100%); HRMS (ESI⁺) C₁₂H₁₈NO₂⁺ ([M + H]⁺) requires 208.1332; found 208.1331. *Method B (from* **34**).²³ OSO₄ (37 mg, 0.14 mmol) was added to a

Method B (from 34).²⁵ OsO₄ (37 mg, 0.14 mmol) was added to a stirred solution of 34 (500 mg, 2.89 mmol) in THF (20 mL) and H₂O (5 mL), followed by a solution of NMO (1.22 g, 10.4 mmol) in H₂O (2.5 mL). The reaction mixture was stirred at rt for 16 h. Saturated aq Na₂SO₃ (1 mL) was then added, and the resultant mixture was allowed to stir at rt for 1 h. The reaction mixture was then extracted with EtOAc (3 × 10 mL), and the combined organic extracts were dried and concentrated in vacuo. Purification via flash column chromatography (CH₂Cl₂/MeOH, 24:1) gave 33 as a yellow oil (399 mg, 67%, >99:1 dr).

N(1)-Benzyl-1,2,3,6-tetrahydropyridine 34. A mixture of pyridine (10.2 mL, 126 mmol) and BnCl (17.5 mL, 152 mmol) was stirred at 140 °C for 1 h, before being allowed to cool to rt. The red resin was dissolved in EtOH (350 mL). Since the resin was barely soluble in EtOH, this dissolution was best performed by repeated addition of portions (~50 mL) of EtOH to the resin, equilibrating the mixture with ultrasonication and decanting the liquid phase. NaBH₄ (10.5 g, 277 mmol) was then added portionwise to the ethanolic solution at 0 °C, and the resultant mixture was allowed to stir at rt for 16 h. H₂O (150 mL) was added, and the organic layer was decanted off from the resultant colorless solid. The solid residue $(2 \times 50 \text{ mL})$ and the aqueous layer $(3 \times 10 \text{ mL})$ were both extracted with Et₂O, and the combined organic extracts were then dried and concentrated in vacuo to give 34 as a yellow oil (17.6 g, 81%);²² $\delta_{\rm H}$ (400 MHz, CDCl₃) 2.18-2.24 (2H, m, C(3)H₂), 2.60 (2H, t, J 5.7, C(2)H₂), 3.01 (2H, m, C(6)H₂), 3.62 (2H, s, NCH₂Ph), 5.66-5.74 (1H, m, C(4)H), 5.76-5.82 (1H, m, C(5)H), 7.26-7.43 (5H, m, Ph).

(RS,SR)-N(1)-Benzyl-3,4-epoxypiperidine 35. HBF₄ (40% aq, 4.53 mL, 28.9 mmol) was added to a stirred solution of 34 (1.00 g, 5.78 mmol) in CH₂Cl₂ (30 mL), and the resultant mixture was allowed to stir at rt for 5 min. m-CPBA (62%, 2.41 g, 8.67 mmol) was then added, and the reaction mixture was stirred at rt for 16 h. Saturated aq Na₂SO₃ (1 mL) was then added until starch-iodide paper indicated that no oxidant remained. The organic layer was washed with NaHCO3 $(3 \times 10 \text{ mL})$, and the combined aqueous layers were extracted with CH_2Cl_2 (3 × 10 mL). The combined organic extracts were then dried and concentrated in vacuo. Purification via flash column chromatography (eluent 30-40 °C petrol/EtOAc, 3:2) gave 35 as a yellow oil (77 mg, 70%);³⁶ δ_{H} (400 MHz, CDCl₃) 1.95–2.09 (2H, m, C(5)H₂), 2.21 (1H, ddd, J 11.5, 9.2, 4.3, C(6)H_A), 2.32-2.38 (1H, m, C(6)H_B), 2.69 (1H, d, J 13.4, $C(2)H_A$), 3.04 (1H, ddd, J 13.4, 4.3, 1.1, $C(2)H_B$), 3.22–3.27 (2H, m, C(3)H, C(4)H), 3.46 (1H, d, J 13.6, NCH_AH_BPh), 3.48 (1H, d, J 13.6, NCH_AH_BPh), 7.24–7.36 (5H, m, Ph).

(*RS,RS*)-*N*(1)-Benzyl-3,4-dihydroxypiperidine 36. Concentrated H₂SO₄ (1.4 mL) and a few drops of H₂O were added to a stirred solution of 35 (1.00 g, 5.28 mmol) in 1,4-dioxane (20 mL), and the resultant mixture was stirred at rt for 16 h and then concentrated in vacuo. NaHCO₃ (5 mL) was added to the residue, and the reaction mixture was extracted with CHCl₃/^jPrOH (3:1, 3 × 10 mL). The combined organic extracts were washed with 2.0 M aq KOH (15 mL), dried, and concentrated in vacuo to give 36 as a yellow oil (874 mg, 80%, >99:1 dr);²¹ $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.57–1.67 (1H, m, C(5)H_A), 1.91–2.00 (1H, m, C(5)H_B), 2.05 (1H, app t, J 9.8, C(2)H_A), 2.15 (1H, app td, J 11.0, 2.7, C(6)H_A), 2.25 (2H, br s, OH), 2.72–2.80 (1H, m, C(6)H_B), 2.95 (1H, dd, J 11.0, 2.7, C(2)H_B),

3.44-3.51 (1H, m, C(4)H), 3.52-3.60 (3H, m, C(3)H, NCH₂Ph), 7.25-7.35 (5H, m, Ph).

(RS,RS)-N(1)-Benzyl-3,4-dihydroxy-3,4-O-carbonylpiperidine 37. CDI (156 mg, 0.96 mmol) and DMAP (12 mg, 98 µmol) were added to a stirred solution of 36 (100 mg, 0.48 mmol, >99:1 dr) in THF (4 mL), and the resultant mixture was allowed to stir at rt for 4 days. NH₄Cl (1 mL) was added, and the reaction mixture was extracted with EtOAc (2×10 mL). The combined organic extracts were then washed with brine (10 mL), dried, and concentrated in vacuo to give 37 as a yellow oil (129 mg, >99:1 dr);³⁷ ν_{max} (ATR) 2925 (C–H), 1815 (C=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.91–2.02 (1H, m, C(5) H_A), 2.25–2.40 (3H, m, C(2) H_A , C(5) H_B , C(6) H_A), 2.92–2.97 (1H, m, C(6)H_B), 3.27 (1H, dd, J 12.5, 4.7, C(2)H_B), 3.63 (2H, s, NCH₂Ph), 5.08-5.15 (1H, m, C(4)H), 5.30 (1H, td, J 9.3, 4.7, C(3)H), 7.27–7.40 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 29.1 (C(5)), 50.1 (*C*(6)), 54.4 (*C*(2)), 61.8 (NCH₂Ph), 74.5 (*C*(3)), 76.5 (*C*(4)), 128.5, 128.8, 130.9 (o,m,p-Ph), 137.1 (i-Ph), 147.9 (CO); m/z (ESI⁺) 208 ([(M-CO)+3H]⁺, 100%); HRMS (TOF MS EI⁺) C₁₃H₁₅NO₃⁺ (M⁺) requires 233.1052; found 233.1052.

(RS,SR)-N(1)-Benzyl-3-iodopiperidin-4-ol 38. I₂ (398 mg, 1.57 mmol) and NaHCO3 (132 mg, 1.57 mmol) were added to a stirred solution of 19 (100 mg, 0.52 mmol) in MeCN (4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was then diluted with Et₂O (15 mL) and washed with satd aq $Na_2S_2O_3$ (15 mL); then the organic layer was dried and concentrated in vacuo to give a 70:30 mixture of 38 and 39, respectively. Purification via flash column chromatography (eluent 30-40 °C petrol/EtOAc, 2:1) gave **38** as a brown solid (40 mg, 24%, >99:1 dr); mp 103–105 °C; ν_{max} (ATR) 3350 (O–H), 2945 (C–H); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.93–2.00 $(1H, m, C(5)H_A)$, 2.08 (1H, app br s, $C(5)H_B$), 2.51 (1H, app br s, $C(6)H_A$, 2.60–2.78 (2H, m, $C(2)H_A$, $C(6)H_B$), 2.92 (1H, app t, J 10.0, C(2)H_B), 3.53 (1H, d, J 13.2, NCH_AH_BPh), 3.61 (1H, d, J 13.2, NCH_AH_BPh), 4.57 (1H, dt, J 8.5, 3.2, C(3)H), 7.24–7.36 (5H, m, *Ph*);³⁸ $\delta_{\rm H}$ (400 MHz, PhMe- d_{8} , 363 K) 1.50–1.59 (1H, m, C(5) $H_{\rm A}$), 1.66-1.73 (1H, m, C(5)H_B), 2.14-2.21 (1H, m, C(6)H_A), 2.41-2.48 $(2H, m, C(2)H_A, C(6)H_B), 2.78$ (1H, dd, J 11.7, 8.8, C(2)H_B), 3.07 (1H, app br s, C(4)H), 3.22 (1H, d, J 13.2, NCH_AH_BPh), 3.31 (1H, d, J 13.2, NCH_AH_BPh), 4.20 (1H, dt, J 8.8, 3.0, C(3)H), 7.00–7.24 (5H, m, Ph); δ_{C} (100 MHz, CDCl₃) 31.9 (C(5)), 39.9 (C(3)), 48.3 (C(6)), 57.4 (*C*(2)), 62.1 (NCH₂Ph), 68.7 (*C*(4)), 127.3, 128.4, 129.0 (*o*,*m*,*p*-Ph), 137.7 (*i*-Ph); ³⁹ $\delta_{\rm C}$ (100 MHz, PhMe- d_8 , 363 K) 32.0 (*C*(5)), 39.7 (C(3)), 48.0 (C(6)), 57.7 (C(2)), 61.8 (NCH_2Ph) , 68.4 (C(4)), 124.4, 124.6, 124.8 (*o*,*m*,*p*-*Ph*), 137.1 (*i*-*Ph*); m/z (ESI⁺) 318 ([M + H]⁺, 100%); HRMS (ESI^+) $C_{12}H_{17}INO^+$ $([M + H]^+)$ requires 318.0349; found 318.0349. Further elution gave a 30:70 mixture of 38 and 39, respectively, as a yellow oil (19 mg, 12%). Data for mixture: $\nu_{\rm max}$ (ATR) 3350 (O-H), 2945 (C-H); m/z (ESI⁺) 318 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{12}H_{17}INO^+$ ([M + H]⁺) requires 318.0349; found 318.0347. Data for 39: $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.71 (1H, dtd, J 12.6, 10.8, 4.3, C(5)H_A), 2.05 (1H, ddt, J 12.6, 4.3, 2.7, C(5)H_B), 2.21 (1H, app td, J 12.6, 2.7, C(6)H_A), 2.56 (1H, t, J 11.5, C(2)H_A), 2.93-2.99 (1H, m, C(6)H_B), 3.28 (1H, app dq, J 11.5, 2.2, C(2)H_B), 3.56 (2H, app d, J 1.7, NCH₂Ph), 3.64 (1H, app td, J 10.8, 4.3, C(4)H), 4.11 (1H, ddd, J 11.5, 9.8, 4.3, C(3)H), 7.25-7.38 (5H, m, Ph); δ_C (100 MHz, CDCl₃) 33.3 (C(5)), 38.4 (C(3)), 51.5 (C(6)), 61.4 (C(2)), 61.4 (NCH₂Ph), 75.2 (C(4)), 127.3 (p-Ph), 128.4, 128.9 (o,m-Ph), 137.8 (i-Ph).

(*RS,RS*)-*N*(1)-Benzyl-3-iodopiperidin-4-ol 39. Step 1. I₂ (752 mg, 2.31 mmol) and NaHCO₃ (586 mg, 2.31 mmol) were added to a stirred solution of 19 (147 mg, 0.77 mmol) in MeCN (4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was then diluted with Et_2O (15 mL) and washed with satd aq Na₂S₂O₃ (15 mL); then the organic layer was dried and concentrated in vacuo to give a 70:30 mixture of 38 and 39, respectively, as a yellow oil (205 mg).

Step 2. NaHCO₃ (586 mg, 2.31 mmol) was added to a stirred solution of the residue of **38** and **39** (70:30, 205 mg) in dioxane/H₂O (3:1, 4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was then diluted with Et₂O (15 mL), and the organic layer was washed with satd aq NaHCO₃ (15 mL), dried, and concentrated in vacuo to give a 70:30 mixture of **32** and **39**, respectively.

Purification via flash column chromatography (eluent 30-40 °C petrol/ acetone, 5:1) gave **39** as a yellow oil (13 mg, 5%, >99:1 dr). Further elution gave **32** as a yellow oil (51 mg, 28%).

(1*RS*,4*SR*,5*SR*)-*N*(1)-Benzyl-4-hydroxy-1-azabicyclo[3.1.0]hexanium Tetrafluoroborate 40. AgBF₄ (18 mg, 95 μmol) was added to a solution of 38 (25 mg, 78 μmol, >99:1 dr) in CD₂Cl₂, and the reaction mixture was shaken at rt for 5 min to give 40 (>99:1 dr); $\delta_{\rm H}$ (400 MHz, CD₂Cl₂) 1.66–1.77 (1H, m, C(3)H_A), 2.42–2.47 (1H, m, C(3)H_B), 3.15 (1H, dd, *J* 7.7, 4.1, C(6)H_A), 3.18–3.22 (1H, m, C(6)H_B), 3.58 (1H, app d, *J* 12.6, C(2)H_A), 3.63 (1H, app d, *J* 12.6, C(2)H_B), 3.96 (1H, app dt, *J* 7.7, 5.5, C(5)H), 4.35 (1H, d, *J* 13.4, NCH_AH_BPh), 4.58 (1H, d, *J* 13.4, NCH_AH_BPh), 5.12 (1H, app td, *J* 8.2, 5.0, C(4)H), 7.19–7.52 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CD₂Cl₂) 28.6 (C(3)), 36.4 (C(6)), 52.0 (C(5)), 54.8 (C(2)), 62.6 (NCH₂Ph), 68.7 (C(4)), 130.3 (*p*-Ph), 128.9, 131.2 (*o*,*m*-Ph), 134.0 (*i*-Ph).

(1*RS*,4*RS*,5*SR*)-*N*(1)-Benzyl-4-hydroxy-1-azabicyclo[3.1.0]hexanium Tetrafluoroborate 41. AgBF₄ (18 mg, 95 μmol) was added to a solution of 39 (25 mg, 78 μmol, >99:1 dr) in CD₂Cl₂, and the reaction mixture was shaken at rt for 5 min to give 41 (>99:1 dr); $\delta_{\rm H}$ (400 MHz, CD₂Cl₂) 1.99–2.08 (1H, m, C(3)H_A), 2.17 (1H, app dd, *J* 15.3, 7.9, C(3)H_B), 2.83 (1H, dd, *J* 6.1, 4.9, C(6)H_A), 3.23 (1H, dd, *J* 8.1, 4.9, C(6)H_B), 3.49–3.57 (1H, m, C(2)H_A), 3.62–3.70 (1H, m, C(2)H_B), 3.92 (1H, app t, *J* 7.1, C(5)H), 4.48 (1H, d, *J* 13.4, NCH_AH_BPh), 4.61 (1H, d, *J* 13.4, NCH_AH_BPh), 4.70 (1H, app d, *J* 4.9, C(4)H), 7.25–7.51 (SH, m, Ph); $\delta_{\rm C}$ (100 MHz, CD₂Cl₂) 30.8 (C(3)), 38.1 (C(6)), 53.7 (C(2)), 57.8 (C(5)), 62.4 (NCH₂Ph), 70.4 (C(4)), 128.9, 129.5, 130.2 (*o*,*m*,*p*-Ph), 131.2 (*i*-Ph).

(*RS,SR*)-*N*(1)-Benzyl-2-hydroxymethyl-3-hydroxypyrrolidine 42. AgBF₄ (18 mg, 95 μ mol) was added to a solution of 39 (25 mg, 78 μ mol, >99:1 dr) in CD₂Cl₂, and the reaction mixture was shaken at rt for 5 min. The reaction mixture was then poured into a stirred solution of NaHCO₃ (20 mg, 0.23 mmol) in dioxane/H₂O (3:1, 4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was then diluted with Et₂O (10 mL), and the organic layer was washed with satd aq NaHCO₃ (10 mL), dried, and concentrated in vacuo to give a 77:15:8 mixture of 42, 36, and 32, respectively (6 mg). Data for 42:⁴⁰ $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.71 (1H, ddt, *J* 13.4, 6.8, 1.9, C(4)H_A), 1.93–2.01 (1H, m, C(4)H_B), 2.61–2.66 (2H, m, C(2)H, C(5)H_A), 2.96–3.01 (1H, m, C(2)CH₂), 3.96 (1H, d, *J* 12.9, NCH_AH_BPh), 3.62–3.65 (2H, m, C(2)CH₂), 3.96 (1H, d, *J* 12.9, NCH_AH_BPh), 4.34 (1H, app dt, *J* 6.8, 2.8, C(3)H), 7.25–7.40 (5H, m, *Ph*).

(RS,SR)-N(1)-Benzyl-3-mercapto-4-hydroxy-3,4-S,O-iminopiperidine 44 and (RS,SR)-(5-N-Benzyl)hexahydro-[1,3]oxathiino-[5,6-b]pyrrol-2-imine 45. AgSCN (44 mg, 0.26 mmol) was added to a stirred solution of 38 (70 mg, 0.22 mmol, >99:1 dr) in CH₂Cl₂ (4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was filtrated through Celite (eluent CH₂Cl₂), and the resultant solution was washed with 2.0 M aq KOH (20 mL) and brine (20 mL), dried, and concentrated in vacuo to give a 85:15 mixture of 44 and 45, respectively. Purification via flash column chromatography (eluent 30-40 °C petrol/EtOAc, 1:4) gave 45 as a yellow oil (4 mg, 4%, >99:1 dr); $\nu_{\rm max}$ (ATR) 3437 (N–H), 1639 (C=N); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.70-1.78 (1H, m, C(7)H_A), 2.19-2.31 (2H, m, $C(6)H_A$, $C(7)H_B$), 2.91 (1H, q, J 6.0, C(4a)H), 3.02–3.07 (1H, m, $C(6)H_B$), 3.24–3.27 (2H, m, $C(4)H_2$), 3.42 (1H, d, J 13.1, NCH_AH_BPh), 3.97 (1H, d, J 13.1, NCH_AH_BPh), 4.42-4.46 (1H, m, C(7a)*H*), 7.28–7.36 (5H, m, *Ph*); δ_{C} (100 MHz, CDCl₃) 33.4 (*C*(7)), 33.7 (C(4)), 51.3 (C(6)), 58.7 (NCH₂Ph), 66.2 (C(4a)), 72.4 (C(7a)), 113.6 (C(2)), 127.4, 128.5, 128.8 (o,m,p-Ph), 137.9 (i-Ph), 169.3 (SCN); m/z (ESI⁺) 249 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{13}H_{17}N_2OS^+$ ([M + H]⁺) requires 249.1056; found 249.1054. Further elution gave 44 as a yellow oil (40 mg, 43%, >99:1 dr); $\nu_{\rm max}$ (ATR) 3300 (N–H), 1639 (C=N); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.94–2.02 (1H, m, C(5) H_A), 2.18–2.33 (3H, m, C(2) H_A , C(5) H_B , $C(6)H_A$, 2.67–2.73 (1H, m, $C(6)H_B$), 3.00 (1H, ddd, J 11.9, 5.8, 1.7, $C(2)H_B$, 3.50 (1H, d, J 13.1, NCH_AH_BPh), 3.57 (1H, d, J 13.1, NCH_AH_BPh), 3.67 (1H, ddd, J 10.8, 5.8, 4.1, C(3)H), 4.64–4.67 (1H, m, C(4)H), 7.26–7.36 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 27.9 (C(5)), 47.1 (C(3)), 47.5 (C(6)), 56.4 (C(2)), 62.5 (NCH₂Ph),

80.3 (*C*(4)), 127.3, 128.3, 129.0 (*o*,*m*,*p*-*Ph*), 137.6 (*i*-*Ph*), 169.1 (SCN); *m*/*z* (ESI⁺) 249 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{13}H_{17}N_2OS^+$ ([M + H]⁺) requires 249.1056; found 249.1054.

(RS,SR)-N(1)-Benzyl-3-mercapto-4-hydroxy-3,4-S,O-carbonylpiperidine 46. A solution of 44 (33 mg, 0.13 mmol) in 1.0 M aq HCl (4 mL) was stirred at rt for 16 h before being concentrated in vacuo. The reaction mixture was then partitioned between H₂O (10 mL) and CHCl₃/ⁱPrOH (3:1, 10 mL). The aqueous layer was extracted with $\text{CHCl}_3\text{/}^i\text{PrOH}$ (3:1, 4 \times 5 mL), and the combined organic extracts were washed with 2.0 M aq KOH, dried, and concentrated in vacuo to give **46** as a yellow oil (13 mg, 39%, >99:1 dr); ν_{max} (ATR) 1732 (C=O); δ_{H} (400 MHz, CDCl₃) 1.96–2.06 (1H, m, $C(5)H_A$), 2.27–2.35 (3H, m, $C(2)H_A$, $C(5)H_B$, $C(6)H_A$), 2.68–2.73 $(1H, m, C(6)H_B)$, 3.05 (1H, ddd, J 12.0, 5.7, 1.8, C(2)H_B), 3.51 (1H, d, J 13.1, NCH_AH_BPh), 3.58 (1H, d, J 13.1, NCH_AH_BPh), 3.78 (1H, ddd, J 10.6, 5.7, 4.4, C(3)H), 4.69-4.73 (1H, m, C(4)H), 7.27-7.37 (5H, m, Ph); δ_{C} (100 MHz, CDCl₃) 27.7 (C(5)), 46.5 (C(3)), 47.5 (C(6)), 56.7 (C(2)), 62.5 (NCH₂Ph), 78.6 (C(4)), 127.4 (p-Ph), 128.4, 128.9 (o,m-Ph), 137.5 (i-Ph), 172.5 (SCO); m/z (ESI⁺) 250 $([M + H]^+, 100\%);$ HRMS (ESI⁺) $C_{13}H_{16}NO_2S^+$ ($[M + H]^+$) requires 250.0896: found 250.0894

(RS,SR)-N(1)-Benzyl-3-amino-4-hydroxy-3,4-N,O-carbonylpiperidine 47 and (RS,RS)-(5-N-Benzyl)hexahydropyrrolo[2,3e][1,3]oxazin-2(3H)-one 48. AgOCN (50 mg, 0.33 mmol) was added to a stirred solution of 38 (88 mg, 0.28 mmol, >99:1 dr) in CH₂Cl₂ (4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was filtrated through Celite (eluent CH_2Cl_2), and the resultant solution was washed with 2.0 M aq KOH (20 mL) and brine (20 mL), dried, and concentrated in vacuo to give an 85:15 mixture of 47 and 48, respectively. Purification via flash column chromatography (eluent 30-40 °C petrol/EtOAc/Et₃N, 80:19:1) gave 47 as a yellow oil (34 mg, 53%, >99:1 dr); ν_{max} (ATR) 1749 (C=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 2.00 (1H, ddd, J 15.5, 10.5, 5.2, C(5)H_A), 2.12-2.20 (2H, m, C(2) H_{A} , C(5) H_{B}), 2.37 (1H, td, J 10.5, 3.5, C(6) H_{A}), 2.58–2.64 (1H, m, C(6)H_B), 2.88 (1H, ddd, J 11.3, 5.8, 1.6, C(2)H_B), 3.50 (1H, d, J 13.2, NCH_AH_BPh), 3.56 (1H, d, J 13.2, NCH_AH_BPh), 3.77 (1H, dt, J 8.2, 5.8, C(3)H), 4.64 (1H, ddd, J 5.8, 4.7, 3.5, C(4)H), 5.41 (1H, br s, NH), 7.26–7.36 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 26.9 (C(5)), 48.1 (C(6)), 51.1 (C(3)), 55.7 (C(2)), 62.4 (NCH₂Ph), 74.3 (C(4)), 127.3 (p-Ph), 128.4, 128.9 (o,m-Ph), 137.7 (i-Ph), 159.9 (NCO); m/z (ESI⁺) 233 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{13}H_{17}N_2O_2^+$ ([M + H]⁺) requires 233.1285; found 233.1282. Further elution gave 48 as a yellow oil (5 mg, 7%, >99:1 dr); ν_{max} (ATR) 1756 (C=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 2.00–2.09 (1H, m, C(7)H_A), 2.15– 2.24 (1H, m, $C(7)H_B$), 2.30–2.37 (1H, m, $C(6)H_A$), 2.83–2.87 (1H, m, C(4a)H), 3.17 (1H, dd, J 8.5, 3.8, C(6)H_B), 3.25 (1H, ddd, J 12.7, 3.8, 3.2, C(4)H_A), 3.35 (1H, ddd, J 12.7, 5.3, 1.4, C(4)H_B), 3.52 (1H, d, J 13.4, NCH_AH_BPh), 3.88 (1H, d, J 13.4, NCH_AH_BPh), 4.82–4.88 (1H, m, C(7a)H), 5.44 (1H, br s, NH), 7.25-7.37 (5H, m, Ph); $\delta_{\rm C}$ (100 MHz, CDCl₃) 31.4 (C(7)), 40.4 (C(4)), 51.5 (C(6)), 57.9 (NCH₂Ph), 59.9 (C(4a)), 80.7 (C(7a)), 127.3, 128.4, 128.4 (*o*,*m*,*p*-Ph), 138.4 (*i-Ph*), 154.4 (C(2)); m/z (ESI⁺) 233 ($[M + H]^+$, 100%); HRMS (ESI⁺) $C_{13}H_{17}N_2O_2^+$ ([M + H]⁺) requires 233.1285; found 233.1282.

(RS,SR)-N(1)-Benzyl-3-(N-benzoylamino)-4-hydroxy-3,4-N,Ocarbonylpiperidine 55. BzNCO (10 μ L, 78 μ mol) was added dropwise to a stirred solution of 38 (25 mg, 78 μ mol, >99:1 dr) in CH₂Cl₂ (2 mL) at rt, and the resultant mixture was stirred at rt for 5 min. AgBF₄ (18 mg, 94 μ mol) was then added in one portion, and the reaction mixture was stirred at rt for 16 h before being filtrated through Celite (eluent CH₂Cl₂). The resultant solution was washed with 2.0 M aq KOH (10 mL) and brine (10 mL), dried, and concentrated in vacuo. Purification via flash column chromatography (eluent 30-40 °C petrol/EtOAc, 2:1) gave 55 as a yellow oil (6 mg, 23%, >99:1 dr); $\nu_{\rm max}$ (ATR) 1786, 1680 (C=O); $\delta_{\rm H}$ (400 MHz, $CDCl_3$) 2.04–2.11 (1H, m, $C(5)H_A$), 2.21 (1H, dq, J 15.1, 3.3, $C(5)H_B$, 2.33–2.41 (2H, m, $C(2)H_A$, $C(6)H_A$), 2.64 (1H, m, C(6)H_B), 3.50 (1H, d, J 13.1, NCH_AH_BPh), 3.51-3.53 (1H, m, $C(2)H_B$, 3.66 (1H, d, J 13.1, NCH_AH_BPh), 4.59 (1H, dt, J 8.2, 5.9, C(3)H), 4.73-4.77 (1H, m, C(4)H), 7.26-7.34 (5H, m, Ph), 7.42-7.45 (2H, m, Ph), 7.54–7.57 (1H, m, Ph), 7.63–7.65 (2H, m, Ph);

$$\begin{split} &\delta_{\rm C} \ (100 \ {\rm MHz}, \ {\rm CDCl}_3) \ 26.8 \ (C(5)), \ 47.2 \ (C(6)), \ 53.7 \ (C(2)), \\ &54.7 \ (C(3)), \ 62.4 \ ({\rm NCH}_2{\rm Ph}), \ 72.8 \ (C(4)), \ 127.3 \ (p\mbox{-}p\mbox{-}h), \ 127.9, \\ &128.4, \ 128.9, \ 129.0 \ (o,m\mbox{-}p\mbox{-}h), \ 132.4 \ (p\mbox{-}p\mbox{-}h), \ 132.9, \ 137.6 \ (i\mbox{-}p\mbox{-}h), \ 153.3 \\ &({\rm NCO}), \ 169.6 \ ({\rm NCOPh}); \ m/z \ ({\rm ESI}^+) \ 337 \ ([{\rm M} + {\rm H}]^+, \ 100\%); \ {\rm HRMS} \\ &({\rm ESI}^+) \ C_{20}{\rm H}_{21}{\rm N}_2{\rm O}_3^+ \ ([{\rm M} + {\rm H}]^+) \ {\rm requires} \ 337.1547; \ {\rm found} \ 337.1537. \end{split}$$

(RS,SR)-N(1)-Benzyl-3-(N-tosylamino)-4-hydroxy-3,4-N,Ocarbonylpiperidine 56. Method A (from 38). TsNCO (22 µL, 145 μ mol) was added dropwise to a stirred solution of 38 (46 mg, 145 μ mol, >99:1 dr) in CH₂Cl₂ (2 mL) at rt, and the resultant mixture was stirred at rt for 5 min. AgBF₄ (37 mg, 188 μ mol) was then added in one portion, and the reaction mixture was stirred at rt for 16 h and filtrated through Celite (eluent CH2Cl2). The resultant solution was washed with 2.0 M aq KOH (20 mL) and brine (20 mL), dried, and concentrated in vacuo to give 56 as a yellow solid (35 mg, 62%, >99:1 dr); mp 111–113 °C; ν_{max} (ATR) 1782 (C=O); δ_{H} (400 MHz, $CDCl_3$) 1.90–2.00 (1H, m, $C(5)H_A$), 2.05–2.18 (3H, m, $C(2)H_{A}$, $C(5)H_{B}$, $C(6)H_{A}$), 2.47 (3H, s, ArMe), 2.56–2.65 (1H, m, C(6)*H*_B), 3.31 (1H, ddd, *J* 11.6, 6.3, 1.8, C(2)*H*_B), 3.47 (1H, d, *J* 13.5, NCH_AH_BPh), 3.50 (1H, d, J 13.5, NCH_AH_BPh), 4.51 (1H, dt, J 9.1, 6.3, C(3)H), 4.60-4.64 (1H, m, C(4)H), 7.21-7.38 (7H, m, Ar, Ph), 7.92 (2H, d, J 8.4, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 21.7 (ArMe), 26.5 (C(5)), 46.8 (C(6)), 54.5 (C(2)), 56.3 (C(3)), 62.4 (NCH₂Ph), 73.3 (C(4)), 127.5 (p-Ph), 128.4, 128.5, 128.9, 129.7 (C(2'), C(3')), C(5'), C(6'), o,m-Ph), 135.4 (C(1')), 137.1 (*i*-Ph), 145.5 (C(4')), 151.9 (NCO) m/z (ESI⁺) 387 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{20}H_{23}N_2O_4S^+$ ([M + H]⁺) requires 387.1373; found 387.1373.

Method B (from 47). Step 1: AgOCN (28 mg, 0.19 mmol) was added to a stirred solution of 38 (50 mg, 0.16 mmol, >99:1 dr) in CH_2Cl_2 (2 mL) at rt, and the resultant mixture was stirred at rt for 16 h. The reaction mixture was filtrated through Celite (eluent CH_2Cl_2), and the resultant solution was washed with 2.0 M aq KOH (15 mL) and brine (15 mL), dried, and concentrated in vacuo to give an 85:15 mixture of 47 and 48, respectively, as a yellow oil (33 mg).

Method B (from 47). Step 2: A solution of the residue of 47 and 48 (85:15, 33 mg) in THF (1 mL) and TsCl (39 mg, 0.20 mmol) was added to a stirred slurry of NaH (60% dispersion in mineral oil, 13 mg, 0.31 mmol) in THF/DMF (1:1, 2 mL) at 0 °C. The resultant mixture was allowed to warm to rt over 16 h. Saturated aq NH₄Cl (1 mL) was then added at -78 °C, and the reaction mixture was allowed to warm to rt. The resultant mixture was partitioned between H₂O (10 mL) and EtOAc (10 mL). The aqueous layer was extracted with EtOAc (2 × 5 mL), and the combined organic extracts were then washed with satd aq NaHCO₃ (5 mL), dried, and concentrated in vacuo to give **56** as a yellow oil (40 mg, >99:1 dr).

(RS,RS)-N(1)-Benzyl-3-(N-tosylamino)-4-hydroxy-3,4-N,Ocarbonylpiperidine 57. TsNCO (22 µL, 148 µmol) was added dropwise to a stirred solution of 39 (47 mg, 148 μ mol, >99:1 dr) in CH₂Cl₂ (2 mL) at rt, and the resultant mixture was stirred at rt for 5 min. AgBF₄ (37 mg, 188 μ mol) was then added in one portion, and the reaction mixture was stirred at rt for 16 h and then filtrated through Celite (eluent CH₂Cl₂). The resultant solution was washed with 2.0 M aq KOH (20 mL) and brine (20 mL), dried, and concentrated in vacuo to give 57 as a yellow oil (17 mg, 30%, >99:1 dr); $\nu_{\rm max}$ (ATR) 1797 (C=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.72 (1H, qd, J 11.5, 4.5, C(5)H_A), 1.96–2.02 (1H, m, C(5)H_B), 2.15 (1H, td, J 11.5, 2.6, C(6)H_A), 2.40 (3H, s, ArMe), 2.47 (1H, t, J 11.5, C(2)H_A), 2.90-2.92 (1H, m, C(6)H_B), 3.48-3.57 (2H, m, C(3)H, NCH_AH_BPh), 3.68-3.75 (2H, m, C(2)H_B, NCH_AH_BPh), 3.82 (1H, td, J 11.5, 4.5, C(4)H), 7.10–7.32 (7H, m, Ar, Ph), 7.81 (2H, d, J 8.2, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 22.7 (ArMe), 27.9 (C(5)), 49.7 (C(6)), 55.4 (C(2)), 61.7 (NCH₂Ph), 62.4 (C(3)), 80.8 (C(4)), 126.2 (p-Ph), 127.5 (C(1')), 128.4, 128.6, 129.9, 132.9 (C(2'), C(3'), C(5'), C(6'), C(6'))o,m-Ph), 137.3 (i-Ph), 145.8 C(4'), 153.1 (NCO); m/z (ESI⁺) 387 $([M + H]^+, 100\%);$ HRMS (ESI⁺) $C_{20}H_{23}N_2O_4S^+$ $([M + H]^+)$ requires 387.1373; found 387.1374.

(*R*,*R*,*R*)-*N*(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3benzyloxy-4-hydroxy-5-(*N*-tosylamino)-4,5-*O*,*N*-carbonylpiperidine 60. *p*-TsNCO (6 μ L, 39 μ mol) was added dropwise to a stirred solution of 12 (20 mg, 32.8 μ mol, >99:1 dr) in CH₂Cl₂ (2 mL) at rt, and the resultant mixture was stirred at rt for 5 min. AgBF₄

(6 mg, 32.8 μ mol) was added in one portion, and the reaction mixture was stirred at rt for 16 h and then filtrated through Celite (eluent CH₂Cl₂). The resultant solution was washed with 2.0 M aq KOH (10 mL) and brine (10 mL), dried, and concentrated in vacuo to give a 55:40:5 mixture of 59, 60, and 61, respectively, as a brown oil (35 mg). Data for 59: $\nu_{\rm max}$ (ATR) 1640 (C=N); $\delta_{\rm H}$ (400 MHz, CDCl₃) 0.98-1.14 (21H, m, Si(CHMe₂)₃), 2.41 (3H, s, ArMe), 2.86-2.94 (3H, m, C(2)H, C(6)H₂), 3.41 (1H, d, J 14.2, NCH_AH_BPh), 3.77-3.83 (1H, m, C(2)CH_AH_B), 3.90 (1H, dd, J 10.4, 4.1, C(2)CH_AH_B), 4.03 (1H, d, J 14.2, NCH_AH_BPh), 4.35–4.37 (1H, m, C(3)H), 4.49 (1H, d, J 11.8, OCH_AH_BPh), 4.72 (1H, d, J 11.8, OCH_AH_BPh), 4.93–4.95 (2H, m, C(4)H, C(5)H), 7.25–7.38 (12H, m, Ar, Ph), 7.89 (2H, d, J 8.5, Ar); m/z (ESI⁺) 679 ([M + H]⁺, 100%). Purification via flash column chromatography (eluent 30-40 °C petrol/Et₂O, 5:1) gave a 90:10 mixture of 60 and 61, respectively, as a yellow oil (10 mg). Data for mixture: ν_{max} (ATR) 1788 (C=O). Data for 60: $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.00–1.11 (21H, m, Si(CHMe₂)₃), 2.38 (3H, s, ArMe), 2.94 (1H, app dd, J 9.5, 4.4, C(2)H), 3.06 (1H, dd, J 13.2, 2.2, C(6)H_A), 3.12 (1H, dd, J 13.2, 3.2, C(6)H_B), 3.50 (1H, app t, J 9.8, C(2)CH_AH_B), 3.57 (1H, d, J 14.3, NCH_AH_BPh), 3.69 (1H, dd, J 10.1, 4.4, C(2)CH_AH_B), 3.75 (1H, d, J 14.3, NCH_AH_BPh), 4.19 (1H, dd, J 3.9, 1.2, C(3)H), 4.46 (1H, d, J 11.7, OCH_AH_BPh), 4.59 (1H, app dt, J 8.8, 3.2, C(5)H), 4.65-4.70 (2H, m, C(4)H, OCH_AH_BPh), 7.12 (2H, dd, J 7.5, 1.8, Ar), 7.23-7.36 (10H, m, Ph), 7.87 (2H, d, J 7.5, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 11.8 (Si(CHMe₂)₃), 18.0 (Si(CHMe₂)₃), 21.6 (ArMe), 50.4 (C(6)), 55.9 $(C(5)), 60.6 (NCH_2Ph), 62.7 (C(2)CH_2), 64.8 (C(2)), 71.8$ (OCH₂Ph), 72.5 (C(3)), 72.9 (C(4)), 127.1, 127.9 (p-Ph), 127.6, 128.2, 128.3, 128.4, 128.5, 129.7 (C(2'), C(3'), C(5'), C(6'), o,m-Ph), 135.3 (*C*(1')), 137.6, 137.9 (*i-Ph*), 145.3 (*C*(4')), 151.8 (NCO); m/z (ESI⁺) 679 ([M + H]⁺, 100%); HRMS (ESI⁺) C₃₇H₅₁N₂O₆SSi⁺ $([M + H]^+)$ requires 679.3232; found 679.3218. Data for 61: $\delta_{\rm H}$ (400 MHz, CDCl₃) 0.99-1.09 (21H, m, Si(CHMe₂)₃), 1.40 (3H, d, J 6.8, $C(\alpha)Me$), 2.43 (3H, s, ArMe), 2.89 (1H, dd, J 13.4, 4.1, $C(6)H_A$, 3.15–3.19 (1H, m, C(2)H), 3.21 (1H, dd, J 13.4, 3.8, $C(6)H_B$, 3.64–3.67 (1H, m, $C(2)CH_AH_B$), 3.83 (1H, dd, J 10.1, 4.1, $C(2)CH_AH_B)$, 4.03 (1H, q, J 6.8, $C(\alpha)H$), 4.17–4.19 (1H, m, C(3)*H*), 4.45–4.49 (1H, m, C(5)*H*), 4.56–4.58 (1H, m, OCH_AH_BPh), 4.58-4.60 (1H, m, C(4)H), 4.69-4.71 (1H, m, OCH_AH_BPh), 7.11-7.13 (2H, m, Ar), 7.23-7.36 (10H, m, Ph), 7.86-7.88 (2H, m, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 9.5 (C(α)Me), 11.5 (Si(CHMe₂)₃), 18.0 $(Si(CHMe_2)_3)$, 21.6 (ArMe), 44.1 (C(6)), 55.3 (C(5)), 58.6 (C(α)), 60.8 (C(2)), 62.9 (C(2)CH₂), 72.2 (OCH₂Ph), 73.1 (C(3)), 73.7 (C(4)), 127.0, 127.3 (p-Ph), 127.6, 128.1, 128.3, 128.4, 128.5, 129.8 (C(2'), C(3'), C(5'), C(6'), o,m-Ph), 135.1 (C(1')), 137.6, 143.1(i-Ph), 145.3 (C(4')), 151.9 (NCO); m/z (ESI^+) 693 $([M + H]^+)$ 100%); HRMS (ESI⁺) $C_{38}H_{53}N_2O_6SSi^+$ ([M + H]⁺) requires 693.3388; found 693.3375. Further elution gave 16 as a colorless oil (8 mg, 48%, >99:1 dr);¹⁵[α]_D²⁰ -32.6 (c 1.0 in CHCl₃); $\delta_{\rm H}$ (400 MHz, CDCl₃) $1.02-1.15 (21H, m, Si(CHMe_2)_3), 2.83 (1H, dd, J 13.7, 1.0, C(6)H_A),$ 2.90 (1H, dd, J 13.7, 2.0, C(6)H_B), 2.98 (1H, app dd, J 9.2, 4.5, C(2)H), 3.43 (1H, d, J 14.2, NCH_AH_BPh), 3.71 (1H, app t, J 9.2, $C(2)CH_{A}H_{B}$, 3.91 (1H, dd, J 10.1, 4.5, $C(2)CH_{A}H_{B}$), 4.06 (1H, d, J 14.2, NCH_AH_BPh), 4.30 (1H, app d, J 3.6, C(3)H), 4.52 (1H, d, J 11.6, OCH_AH_BPh), 4.70–4.78 (2H, m, C(5)H, OCH_AH_BPh), 4.79– 4.84 (1H, dd, J 8.3, 3.6, C(4)H), 7.23-7.40 (10H, m, Ph).

(*R*,*R*,*R*)-*N*(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3benzyloxy-4-hydroxy-5-(*N*-tosylamino)piperidine 64, (2*R*,3*R*,4*R*,5S)-*N*(1)-benzyl-2-[(triisopropylsilyloxy)methyl]-3benzyloxy-4-hydroxy-5-(*N*-tosylamino)piperidine 65, and (*R*,*R*,*R*,*R*)-*N*(1)-(α-methylbenzyl)-2-[(triisopropylsilyloxy)methyl]-3-benzyloxy-4-hydroxy-5-(*N*-tosylamino)piperidine 66. *Step 1*. I₂ (363 mg, 1.43 mmol) and NaHCO₃ (120 mg, 1.43 mmol) were added to a stirred solution of 11¹⁵ (280 mg, 0.48 mmol, >99:1 dr) in MeCN (4 mL) at rt, and the resultant mixture was stirred at rt for 16 h. TsNCO (0.33 mL, 2.16 mmol) was added dropwise, and the reaction mixture was allowed to stir at rt for 16 h. The reaction mixture was diluted with Et₂O (20 mL), washed with satd aq Na₂S₂O₃ (20 mL), dried, and concentrated in vacuo to give a 76:19:5 mixture of 60, 63, and 61, respectively (300 mg).

Step 2. K_2CO_3 (267 mg, 1.93 mmol) was added to a stirred solution of the residue of 60, 63, and 61 (57:14:4:25, 300 mg) in MeOH (5 mL) at rt, and the resultant mixture was allowed to stir at rt for 16 h. The resultant mixture was concentrated in vacuo, and the residue was partitioned between H2O (20 mL) and EtOAc (20 mL). The aqueous layer was extracted with EtOAc (2×10 mL), and the combined organic extracts were then dried and concentrated in vacuo to give a 76:19:5 mixture of 64, 65, and 66, respectively. Purification via flash column chromatography (eluent 30-40 °C petrol/acetone, 9:1) gave 66 as a colorless oil (6 mg, 2% from 11, >99:1 dr); $[\alpha]_{D}^{20}$ +5.0 (c 0.06 in CHCl₃); ν_{max} (ATR) 3532, 3276 (O–H, N–H), 2942, 2865 (C–H), 1160, 1092 (S=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.03-1.12 (21H, m, Si(CHMe₂)₃), 1.22 (3H, d, J 6.8, C(α)Me), 1.96 (1H, dd, J 12.3, 4.3, C(6)H_A), 2.16 (1H, dd, J 12.3, 1.6, C(6)H_B), 2.37 (3H, s, ArMe), 2.59 (1H, ddd, J 8.8, 4.5, 1.3, C(2)H), 2.97 (1H, d, J 8.8, OH), 3.03-3.07 (1H, m, C(5)H), 3.31 (1H, app t, J 8.8, C(3)H), 3.54 (1H, app td, J 8.8, 3.9, C(4)H), 3.99 (1H, dd, J 11.0, 4.5, $C(2)CH_AH_B$, 4.16 (1H, dd, J 11.0, 1.3, $C(2)CH_AH_B$), 4.60 (1H, d, J 11.0, OCH_AH_BPh), 4.65 (1H, q, J 6.8, C(α)H), 5.07–5.13 (2H, m, OCH_AH_BPh, NH), 6.97–6.99 (2H, m, Ar), 7.02–7.05 (2H, m, Ar), 7.27–7.43 (8H, m, Ph), 7.48–7.51 (2H, m, Ph); $\delta_{\rm C}$ (100 MHz, $CDCl_3$) 9.6 ($C(\alpha)Me$), 11.9 ($Si(CHMe_2)_3$), 18.0 ($Si(CHMe_2)_3$), 21.5 (ArMe), 46.1 (C(6)), 52.4 (C(5)), 54.8 $(C(\alpha))$, 63.3 $(C(2)CH_2)$, 64.6 (C(2)), 74.7 (OCH_2Ph) , 75.1 (C(4)), 78.9 (C(3)), 127.6, 127.6 (p-Ph), 127.1, 127.9, 128.1, 128.3, 128.7, 129.6 (C(2'), C(3'), C(5'), $\tilde{C}(6')$, o,m-Ph), 135.7 (C(1')), 138.6 (*i*-Ph), 143.2 (C(4')), 144.2 (i-Ph); m/z (ESI⁺) 667 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{37}H_{55}N_2O_5SSi^+$ ([M + H]⁺) requires 667.3595; found 667.3571. Further elution gave 64 as a colorless oil(78 mg, 25% from 11, >99:1 dr); $[\alpha]_{\rm D}^{20}$ -16.2 (c 0.5 in CHCl₃); $\nu_{\rm max}$ (ATŘ) 3532, 3276 (O-H, N–H), 2942, 2891 (C–H), 1160, 1091 (S=O); δ_H (400 MHz, CDCl₃) 1.02–1.13 (21H, m, Si(CHM e_2)₃), 2.08 (1H, dd, J 12.3, 2.4, C(6) H_A), 2.32 (1H, dd, J 12.3, 5.3, C(6)H_B), 2.36 (3H, s, ArMe), 2.37-2.41 (1H, m, C(2)H), 3.12-3.16 (2H, m, OH, NCH_AH_BPh), 3.27-3.32 (1H, m, C(5)H), 3.39 (1H, app t, J 8.0, C(3)H), 3.57 (1H, app td, J 8.0, 3.9, C(4)H), 3.88 (1H, dd, J 11.0, 4.4, C(2)CH_AH_B), 4.15 (1H, dd, J 11.0, 1.9, C(2)CH_AH_B), 4.39 (1H, d, J 13.2, NCH_AH_BPh), 4.57 (1H, d, J 11.2, OCH_AH_BPh), 4.97 (1H, d, J 11.2, OCH_AH_BPh), 5.22 (1H, d, J 8.4, NH), 7.05 (2H, d, J 8.0, Ar), 7.26–7.42 (12H, m, Ar, Ph); $\delta_{\rm H}$ (400 MHz, MeOH- d_4) 0.96–1.02 (21H, m, Si(CHMe₂)₃), 2.16 (1H, dd, J 12.5, 4.3, C(6)H_A), 2.35 (3H, s, ArMe), 2.67 (1H, dd, J 12.5, 9.5, C(6)H_B), 2.75-2.78 (1H, m, C(2)H), 3.44-3.49 (1H, m, C(5)H), 3.60 (1H, d, J 13.7, NCH_AH_BPh), 3.64 (1H, app t, J 4.0, C(3)H), 3.75 (1H, app t, J 4.0, C(4)H), 3.92 (1H, dd, J 10.1, 6.0, $C(2)CH_AH_B$, 3.99 (1H, dd, J 10.1, 5.8, $C(2)CH_AH_B$), 4.02 (1H, d, J 13.7, NCH_AH_BPh), 4.47 (1H, d, J 12.0, OCH_AH_BPh), 4.58 (1H, d, J 12.0, OCH_AH_BPh), 7.12 (2H, d, J 8.0, Ar), 7.18–7.32 (10H, m, Ph), 7.62 (2H, d, J 8.0, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 11.9 (Si(CHMe₂)₃), 18.0 $(Si(CHMe_2)_3)$, 21.5 (ArMe), 51.8 (C(6)), 52.0 (C(5)), 57.2 (NCH₂Ph), 63.0 (C(2)CH₂), 65.9 (C(2)), 73.6 (C(4)), 74.2 (OCH₂Ph), 78.7 (C(3)), 127.3, 127.6 (*p*-Ph), 127.0, 127.9, 128.3, 128.6, 129.1, 129.6 (C(2'), C(3'), C(5'), C(6'), o,m-Ph), 136.3 (C(1')), 138.4, 139.3 (*i-Ph*), 143.2 (C(4')); δ_{C} (100 MHz, MeOH- d_{4}) 13.3 $(Si(CHMe_2)_3)$, 18.7 $(Si(CHMe_2)_3)$, 21.6 (ArMe), 51.3 (C(6)), 51.8 (C(5)), 60.6 (NCH₂Ph), 63.3 (C(2)CH₂), 64.1 (C(2)), 73.0 (C(4)), 73.3 (OCH₂Ph), 79.0 (C(3)), 127.8, 128.6 (p-Ph), 128.0, 128.9, 129.2, 129.4, 130.1, 130.1 (*C*(2'), *C*(3'), *C*(5'), *C*(6'), *o*,*m*-*Ph*), 140.4, 141.4 (*i-Ph*), 141.7 (C(4')), 144.3 (C(1')); m/z (ESI⁺) 653 $([M + H]^+, 100\%);$ HRMS (ESI^+) $C_{36}H_{53}N_2O_5SSi^+$ $([M + H]^+)$ requires 653.3439; found 653.3411. Further elution gave 65 as a colorless oil (22 mg, 7% from 11, >99:1 dr); $[\alpha]_{\rm D}^{20}$ -5.6 (c 0.5 in CHCl₃); ν_{max} (ATR) 3532, 3276 (O–H, N–H), 2926, 2865 (C–H), 1094, 1158 (S=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.01–1.08 (21H, m, $Si(CHMe_2)_3$, 2.12 (1H, dd, J 12.1, 6.6, C(6)H_A), 2.39 (3H, s, ArMe), 2.70 (1H, app q, J 4.0, C(2)H), 3.04 (1H, dd, J 12.1, 3.5, C(6)H_B), 3.25-3.30 (1H, m, C(5)H), 3.43-3.56 (4H, m, C(3)H, C(4)H, NCH_AH_BPh, OH), 3.76 (1H, dd, J 11.0, 4.0, C(2)CH_AH_B), 4.01 (1H, d, J 13.7, NCH_AH_BPh), 4.16 (1H, dd, J 11.0, 3.2, C(2)CH_AH_B), 4.56 (1H, d, J 11.5, OCH_AH_BPh), 4.66 (1H, d, J 11.5, OCH_AH_BPh), 5.35 (1H, d, J 8.2, NH), 7.13-7.15 (2H, m, Ar), 7.25-7.38 (10H, m, Ph),

7.54–7.58 (2H, m, Ar); $\delta_{\rm H}$ (400 MHz, MeOH- d_4) 0.95–1.02 (21H, m, Si(CHMe₂)₃), 1.71 (1H, app t, J 11.2, C(6)H_A), 2.30 (1H, app dd, J 9.8, 5.2, C(2)H), 2.45 (1H, dd, J 11.2, 4.5, C(6)H_B), 2.81 (1H, app dd, J 9.8, 4.5, C(5)H), 3.07 (1H, d, J 13.6, NCH_AH_BPh), 3.20 (1H, app t, J 9.8, C(3)H), 3.39 (1H, app t, J 9.8, C(4)H), 3.85 (1H, dd, J 11.2, 5.2, C(2)CH_AH_B), 4.17 (1H, app d, J 9.8, C(2)CH_AH_B), 4.38 (1H, d, J 13.6, NCH_AH_BPh), 4.58 (1H, d, J 12.0, OCH_AH_BPh), 5.08 (1H, d, J 12.0, OCH_AH_BPh), 7.00 (2H, d, J 8.0, Ar), 7.18-7.37 (10H, m, Ph), 7.50 (2H, d, J 8.0, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 11.8 (Si(CHMe₂)₃), 18.0 (Si(CHMe₂)₃), 21.5 (ArMe), 50.9 (C(6)), 53.9 (C(5)), 57.5 (NCH₂Ph), 61.9 (C(2)CH₂), 63.0 (C(2)), 71.7 (C(4)), 73.1 (OCH₂Ph), 79.5 (C(3)), 127.0, 127.8 (*p*-Ph), 126.9, 127.7, 128.3, 128.5, 128.5, 129.6 (C(2'), C(3'), C(5'), C(6'), o,m-Ph), 137.7 (C(1')), 138.0, 138.7 (*i-Ph*), 143.1 (C(4')); δ_{C} (100 MHz, MeOH- d_{4}) 13.3 $(Si(CHMe_2)_3)$, 18.7 $(Si(CHMe_2)_3)$, 21.6 (ArMe), 57.0 (C(5)), 57.9 (C(6)), 59.0 (NCH₂Ph), 65.2 (C(2)CH₂), 69.8 (C(2)), 75.7 (OCH₂Ph), 81.4 (C(3)), 81.8 (C(4)), 127.8, 128.6 (p-Ph), 128.0, 128.9, 129.2, 129.4, 130.1, 130.1 (C(2'), C(3'), C(5'), C(6'), o,m-Ph), 140.4, 141.4 (*i-Ph*), 141.7 (C(4')), 144.3 (C(1')); m/z (ESI⁺) 653 $([M + H]^+, 100\%);$ HRMS (ESI^+) $C_{36}H_{53}N_2O_5SSi^+$ $([M + H]^+)$ requires 653.3439; found 653.3413.

(2R, 3R, 4S, 5S)-N(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3benzyloxy-4-hydroxy-5-(N-tosylamino)-4,5-O,N-carbonylpiperidine 68. I_2 (129 mg, 0.51 mmol) and NaHCO₃ (43 mg, 0.51 mmol) were added to a stirred solution of 67^{15} (100 mg, 0.17 mmol, >99:1 dr) in MeCN (2 mL) at rt, and the resultant mixture was stirred at rt for 16 h. TsNCO (0.12 mL, 0.77 mmol) was added dropwise, and the reaction mixture was allowed to stir at rt for 16 h. The reaction mixture was then diluted with Et₂O (15 mL), washed with satd aq Na₂S₂O₃ (15 mL), dried, and concentrated in vacuo. Purification via flash column chromatography (eluent 30-40 °C petrol/EtOAc, 9:1) gave 68 as a yellow oil (56 mg, 48%, >99:1 dr); $[\alpha]_{D}^{20}$ +14.8 (c 1.0 in CHCl₃); ν_{max} (ATR) 2942, 2865 (C–H), 1785 (C=O), 1173, 1100 (S=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 1.02-1.11 (21H, m, Si(CHMe₂)₃), 2.36 (3H, s, ArMe), 2.94 (1H, app quintet, *J* 3.2, C(2)*H*), 3.10 (1H, dd, *J* 12.0, 7.1, C(6)*H*_A), 3.43 (1H, dd, *J* 12.0, 6.0, C(6)H_B), 3.60-3.66 (2H, m, NCH_AH_BPh, C(2)CH_AH_B), 3.70-3.73 (2H, m, NCH_AH_BPh, C(2)CH_AH_B), 4.12 (1H, app t, J 3.2, C(3)*H*), 4.51 (1H, d, *J* 11.4, OCH_AH_BPh), 4.54–4.58 (2H, m, C(5)*H*, OCH_AH_BPh), 4.84 (1H, dd, J 9.1, 3.2, C(4)H), 7.16–7.20 (6H, m, Ar, *Ph*), 7.25–7.33 (6H, m, *Ph*), 7.87 (2H, dd, J 8.4, *Ar*); $\delta_{\rm C}$ (100 MHz, CDCl₃) 11.8 (Si(CHMe₂)₃), 18.1 (Si(CHMe₂)₃), 21.6 (ArMe), 50.6 (C(6)), 55.6 (C(5)), 58.8 (NCH_2Ph) , 61.4 (C(2)), 61.8 $(C(2)CH_2)$, 72.0 (C(4)), 72.5 (OCH₂Ph), 74.5 (C(3)), 127.3, 127.6 (*p*-Ph), 127.6, 128.1, 128.3, 128.3, 128.4, 129.6 (*C*(2'), *C*(3'), *C*(5'), *C*(6'), *o*,*m*-*Ph*), 135.2 (*C*(1')), 137.5, 137.9 (*i*-*Ph*), 145.1 (*C*(4')), 152.5 (NCO); m/z (ESI⁺) 679 ([M + H]⁺, 100%); HRMS (ESI⁺) C₃₇H₅₁N₂O₆SSi⁺ $([M + H]^+)$ requires 679.3232; found 679.3219.

(2R,3R,4S,5S)-N(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3benzyloxy-4-hydroxy-5-(N-tosylamino)piperidine 69. K₂CO₃ (175 mg, 1.26 mmol) was added to a stirred solution of 68 (85 mg, 0.12 mmol, >99:1 dr) in MeOH (4 mL) at rt, and the resultant mixture was allowed to stir at rt for 16 h. The reaction mixture was then concentrated in vacuo, and the residue was partitioned between H₂O (15 mL) and EtOAc (15 mL). The aqueous layer was extracted with EtOAc (2 \times 10 mL), and the combined organic extracts were then dried and concentrated in vacuo to give **69** as a white solid (68 mg, 83%, >99:1 dr); mp 88–90 °C; $[\alpha]_D^{20}$ +6.0 (c 1.0 in CHCl₃); $\nu_{\rm max}$ (ATR) 3532, 3276 (O–H, N–H), 2942, 2865 (C–H), 1169, 1160, 1091 (S=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 0.99–1.09 (21H, m, Si(CHMe₂)₃), 2.27 (1H, dd, J 12.0, 5.8, C(6)H_A), 2.36 (3H, s, ArMe), 2.60 (1H, d, J 7.6, OH), 2.62 (1H, dd, J 12.0, 2.8, C(6)H_B), 2.89-2.92 (1H, m, C(2)H), 3.51-3.57 (2H, m, C(5)H, NCH_AH_BPh), 3.67 (1H, app t, J 4.0, C(3)H), 3.80-3.86 (2H, m, NCH_AH_BPh, $C(2)CH_AH_B$, 3.87–3.91 (2H, m, $C(2)CH_AH_B$, C(4)H), 4.46 (1H, d, J 11.5, OCH_AH_BPh), 4.51 (1H, d, J 11.5, OCH_AH_BPh), 5.63 (1H, d, J 9.9, NH), 7.10 (2H, d, J 8.0, Ar), 7.24-7.39 (10H, m, Ph), 7.53 (2H, d, J 8.0, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 11.8 (Si(CHMe₂)₃), 18.0 $(Si(CHMe_2)_3)$, 21.5 (ArMe), 50.2 (C(6)), 53.0 (C(5)), 57.6 (NCH₂Ph), 59.5 (C(2)CH₂), 60.4 (C(2)), 66.2 (C(4)),

72.0 (OCH₂Ph), 77.6 (C(3)), 127.1, 128.0 (*p*-*Ph*), 126.6, 127.7, 128.3, 128.5, 128.6, 129.5 (C(2'), C(3'), C(5'), C(6'), *o*,*m*-*Ph*), 137.6 (*i*-*Ph*), 138.5 (C(1')), 139.0 (*i*-*Ph*), 142.8 (C(4')); m/z (ESI⁺) 653 ([M + H]⁺, 100%); HRMS (ESI⁺) C₃₆H₅₃N₂O₅SSi⁺ ([M + H]⁺) requires 653.3439; found 653.3436.

(R,R,R,R)-N(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3,4-dihydroxy-5-aminopiperidine 70 and (R,R,R,R)-N(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3,4-dihydroxy-5-(N-tosylamino)piperidine 71. Naphthalene (346 mg, 2.70 mmol) was dissolved in DME (3 mL), then Na (46 mg, 2.02 mmol) was added under nitrogen, and the resultant green solution was stirred at rt for 2 h. A solution of 64 (60 mg, 92 μ mol, >99:1 dr) in DME (3 mL) was added via cannula at -78 °C; then the resultant mixture was allowed to warm gradually to rt and stirred at rt for 16 h. The reaction mixture was then cooled to 0 °C, and H₂O (1 mL) was added. The reaction mixture was then allowed to warm to rt, and Et₂O (15 mL) was added. The organic layer was washed with satd aq NH4Cl (15 mL) and brine (15 mL) before being dried and concentrated in vacuo to give a 95:5 mixture of 70 and 71, respectively. Purification via flash column chromatography (eluent CH₂Cl₂/MeOH/NH₄OH, 95:4:1) gave 71 as a colorless oil (2 mg, 4%, >99:1 dr); $[\alpha]_{D}^{20}$ -44.0 (c 0.2 in CHCl₃); ν_{max} (ATR) 3322 (O-H, N-H), 2940, 2865 (C-H), 1160, 1094, 1070 (S=O); $\delta_{\rm H}$ (400 MHz, MeOH- d_4) 1.08–1.15 (21H, m, Si(CHMe₂)₃), 1.96 (1H, dd, J 12.0, 1.9, C(6)H_A), 2.21 (1H, dd, J 12.0, 4.6, C(6)H_B), 2.32-2.36 (1H, m, C(2)H), 2.34 (3H, s, ArMe), 3.18 (1H, d, J 13.4, NCH_AH_BPh), 3.37-3.40 (2H, m, C(4)H, C(5)H), 3.52-3.55 (1H, m, C(3)H), 4.10 (1H, dd, J 11.0, 4.7, C(2)CH_AH_B), 4.19 (1H, dd, J 11.0, 2.7, C(2)CH_AH_B), 4.23 (1H, d, J 13.4, NCH_AH_BPh), 7.09 (2H, d, J 8.0, Ar), 7.24–7.40 (7H, m, Ar, Ph); $\delta_{\rm C}$ (100 MHz, MeOH- d_4) 13.3 $(Si(CHMe_2)_3)$, 18.7 $(Si(CHMe_2)_3)$, 21.6 (ArMe), 52.2 (C(6)), 53.6 (C(5)), 58.8 (NCH₂Ph), 63.0 (C(2)CH₂), 69.4 (C(2)), 70.9 (C(3)), 74.7 (C(4)), 128.3 (p-Ph), 128.0, 129.7, 130.1, 130.8 (C(2'), C(3'), C(5'), C(6'), o,m-Ph), 139.1 (C(1')), 141.0 (i-Ph), 144.5(C(4')); m/z (ESI⁺) 563 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{29}H_{47}N_2O_5SSi^+$ ([M + H]⁺) requires 563.2969; found 563.2954. Further elution gave 70 as a colorless oil (23 mg, 63%, >99:1 dr); $[\alpha]_{\rm D}^{20}$ –28.4 (*c* 0.5 in MeOH); $\nu_{\rm max}$ (ATR) 3360 (O–H, N–H), 2942, 2865 (С–Н); $\delta_{\rm H}$ (400 MHz, MeOH- d_4) 1.10–1.17 (21H, m, Si(CHMe₂)₃), 2.19–2.24 (2H, m, C(6)H_A, C(2)H), 2.77 (1H, dd, J 12.0, 3.6, C(6)H_B), 2.79–2.83 (1H, m, C(5)H), 3.17 (1H, d, J 13.4, NCH_AH_BPh), 3.41 (1H, dd, J 8.9, 4.0, C(4)H), 3.53 (1H, app t, J 8.9, C(3)*H*), 4.11 (1H, dd, *J* 11.0, 4.3, C(2)*CH*_AH_B), 4.28 (1H, dd, *J* 11.0, 2.1, C(2)CH_AH_B), 4.47 (1H, d, J 13.4, NCH_AH_BPh), 7.20-7.36 (5H, m, Ph); δ_C (100 MHz, MeOH-d₄) 13.4 (Si(CHMe₂)₃), 18.7 $(Si(CHMe_2)_3)$, 51.6 (C(5)), 55.1 (C(6)), 58.7 (NCH_2Ph) , 63.4 $(C(2)CH_2)$, 70.2 (C(3)), 70.3 (C(2)), 76.7 (C(4)), 128.1 (p-Ph), 129.5, 130.1 (*o*,*m*-Ph), 141.2 (*i*-Ph); m/z (ESI⁺) 409 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{22}H_{41}N_2O_3Si^+$ ([M + H]⁺) requires 409.2881; found 409.2872.

(2R,3R,4S,5S)-N(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3,4-dihydroxy-5-aminopiperidine 72 and (2R,3R,4S,5S)-N(1)-Benzyl-2-[(triisopropylsilyloxy)methyl]-3,4-dihydroxy-5-(Ntosylamino)piperidine 73. Naphthalene (461 mg, 3.60 mmol) was dissolved in DME (3 mL), then Na (62 mg, 2.70 mmol) was added under nitrogen, and the resultant green solution was stirred at rt for 2 h. A solution of 69~(79 mg, 0.12 mmol, >99:1 dr) in DME (3 mL) was added via cannula at -78 °C, and the resultant mixture was allowed to warm gradually to rt and stirred at rt for 16 h. The reaction mixture was then cooled to 0 °C, and H₂O (1 mL) was added. The reaction mixture was then allowed to warm to rt, and Et₂O (15 mL) was added. The organic layer was washed with satd aq NH₄Cl (15 mL) and brine (15 mL) before being dried and concentrated in vacuo. Purification via flash column chromatography (eluent CH₂Cl₂/ MeOH/NH₄OH, 95:4:1) gave 73 as a colorless oil (4 mg, 6%, >99:1 dr); $[\alpha]_D^{20}$ -7.1 (c 0.5 in CHCl₃); ν_{max} (ATR) 3463 (O-H, N–H), 2942, 2866 (С–H), 1159, 1092, 1058 (S=O); $\delta_{\rm H}$ (400 MHz, CDCl₃) 0.99–1.12 (21H, m, Si(CHMe₂)₃), 2.35 (1H, dd, J 11.2, 10.2, $C(6)H_A$, 2.39 (3H, s, ArMe), 2.58 (1H, dd, J 11.2, 4.5, $C(6)H_B$), 2.69 (1H, app td, J 7.8, 4.3, C(2)H), 2.73 (1H, br s, OH), 3.40 (1H, d, J 14.0, NCH_AH_BPh), 3.45–3.51 (1H, m, C(5)H), 3.68–3.77 (4H, m,

NCH_AH_BPh, C(2)CH_AH_B, C(3)H, C(4)H), 4.15 (1H, dd, J 9.9, 4.3, C(2)CH_AH_B), 4.49 (1H, br s, OH), 5.27 (1H, br s, NH), 7.16 (2H, d, J 8.0, Ar), 7.20-7.22 (2H, m, Ph), 7.25-7.33 (3H, m, Ph), 7.62 (2H, d, J 8.0, Ar); $\delta_{\rm H}$ (400 MHz, MeOH- d_4) 1.05–1.12 (21H, m, $Si(CHMe_2)_3$, 2.11 (1H, app t, J 11.0, C(6)H_A), 2.16-2.19 (1H, m, $C(6)H_B$, 2.37 (3H, s, ArMe), 2.45–2.50 (1H, m, C(2)H), 3.13 (1H, d, J 13.7, NCH_AH_BPh), 3.25 (1H, ddd, J 11.0, 4.7, 2.7, C(5)H), 3.42 (1H, dd, J 9.5, 3.1, C(3)H), 3.78 (1H, app t, J 3.1, C(4)H), 3.96 (1H, dd, J 11.0, 4.7, C(2)CH_AH_B), 4.23 (1H, dd, J 11.0, 2.1, C(2)CH_AH_B), 4.31 (1H, d, J 13.7, NCH_AH_BPh), 7.13-7.25 (7H, m, Ar, Ph), 7.53-7.58 (2H, m, Ar); $\delta_{\rm C}$ (100 MHz, CDCl₃) 11.6 (Si(CHMe₂)₃), 17.6 $(Si(CHMe_2)_3)$, 21.6 (ArMe), 51.2 (C(6)), 51.6 (C(5)), 57.7 (NCH₂Ph), 60.3 (C(2)), 66.0 (C(2)CH₂), 68.6 (C(3)), 73.6 (C(4)), 127.1 (p-Ph), 126.7, 128.1, 128.4, 129.7 (C(2'), C(3')), $C(5'), C(6'), o,m-Ph), 138.2 (C(1')), 138.8 (i-Ph), 143.2 (C(4')); \delta_{C}$ (100 MHz, MeOH-d₄) 13.3 (Si(CHMe₂)₃), 18.7 (Si(CHMe₂)₃), 21.7 (ArMe), 50.9 (C(6)), 53.7 (C(5)), 58.6 (NCH₂Ph), 64.3 (C(2)CH₂), 64.5 (C(2)), 70.3 (C(3)), 72.1 (C(4)), 128.0 (p-Ph), 127.8, 129.4, 130.0, 130.8 (C(2'), C(3'), C(5'), C(6'), o,m-Ph), 140.2 (C(1')), 140.6 (*i-Ph*), 144.4 (C(4')); m/z (ESI⁺) 563 ($[M + H]^+$, 100%); HRMS (ESI⁺) $C_{29}H_{47}N_2O_5SSi^+$ ([M + H]⁺) requires 563.2969; found 563.2955. Further elution gave 72 as a colorless oil (30 mg, 61%, >99:1 dr); $[\alpha]_{D}^{20}$ -14.7 (c 0.3 in MeOH); ν_{max} (ATR) 3337 (O-H, N–H), 2942, 2866 (C–H); $\delta_{\rm H}$ (400 MHz, MeOH- d_4) 1.09–1.14 (21H, m, Si(CHMe₂)₃), 2.53 (1H, dd, J 11.9, 7.5, C(6)H_A), 2.83-2.89 $(2H, m, C(2)H, C(6)H_B), 3.12-3.15 (1H, m, C(5)H), 3.59 (1H, d, d)$ J 13.4, NCH_AH_BPh), 3.81-3.83 (1H, m, C(3)H), 3.97 (1H, app t, J 3.2, C(4)H), 4.03 (1H, dd, J 10.9, 4.6, C(2)CH_AH_B), 4.13–4.19 (2H, m, C(2)CH_AH_B, NCH_AH_BPh), 7.21–7.25 (1H, m, Ph), 7.29–7.33 (2H, m, Ph), 7.38–7.39 (2H, m, Ph); $\delta_{\rm C}$ (100 MHz, MeOH-d₄) 13.3 $(Si(CHMe_2)_3)$, 18.7 $(Si(CHMe_2)_3)$, 50.3 (C(6)), 52.1 (C(5)), 59.0 (NCH₂Ph), 62.2 (C(2)CH₂), 65.1 (C(2)), 68.6 (C(4)), 70.9 (C(3)), 128.4 (p-Ph), 129.6, 130.1 (o,m-Ph), 140.5 (i-Ph); m/z (ESI⁺) 409 $([M + H]^+, 100\%);$ HRMS (ESI^+) $C_{22}H_{41}N_2O_3Si^+$ $([M + H]^+)$ requires 409.2881; found 409.2874.

(R,R,R,R)-2-Amino-1,2,5-trideoxy-1,5-imino-p-mannose [(-)-2-amino-1-deoxymannojirimycin (ADMJ)] 25. Step 1. A solution of 70 (30 mg) in 6.0 M aq HCl (1 mL) and MeOH (1 mL) was stirred at 40 °C for 16 h before being concentrated in vacuo to give (R,R,R,R)-N(1)-benzyl-2-hydroxymethyl-3,4-dihydroxy-5-aminopiperidine hydrochloride as a yellow oil (20 mg); $[\alpha]_{\rm D}^{20}$ +31.8 (c 1.0 in MeOH); $\nu_{\rm max}$ (ATR) 3350 (O–H, N–H), 2942, 2865 (C–H); $\delta_{\rm H}$ (400 MHz, MeOH-d₄) 3.51-3.58 (2H, m, C(2)H, C(6)H_A), 3.65-3.76 (2H, m, C(2)C H_AH_B , C(6) H_B), 4.00–4.08 (1H, m, C(4)H), 4.09-4.14 (2H, m, C(2)CH_AH_B, C(3)H), 4.25-4.39 (1H, m, C(5)H), 4.54-4.68 (1H, m, NCH_AH_BPh), 5.00-5.09 (1H, m, NCH_AH_BPh), 7.45–7.53 (3H, m, Ph), 7.61–7.71 (2H, m, Ph); $\delta_{\rm C}$ (100 MHz, MeOH-*d*₄) 44.6 (*C*(5)), 45.3 (*C*(6)), 57.9 (*C*(2)*C*H₂), 60.9 (NCH₂Ph), 64.7 (C(2)), 67.6 (C(3)), 69.0 (C(4)), 130.5, 131.3 (o,m-Ph), 132.7 (p-Ph), 132.8 (i-Ph); m/z (ESI⁺) 253 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{13}H_{21}N_2O_3^+$ ([M + H]⁺) requires 253.1547; found 253.1545.

Step 2. $Pd(OH)_2/C$ (10 mg) was added to a stirred solution of the residue of (R,R,R,R)-N(1)-benzyl-2-hydroxymethyl-3,4-dihydroxy-5aminopiperidine hydrochloride (20 mg) in degassed MeOH (2 mL), and the resultant suspension was stirred at rt for 48 h under an atmosphere of H₂ (5 atm). HCl (1.0 M in Et₂O, 1 mL) was then added, and the resultant suspension was stirred for a further 5 min before being filtrated through Celite (eluent MeOH) and concentrated in vacuo. Purification via ion exchange chromatography on Dowex-50WX8 resin (hydrogen form, 100-200 mesh, eluent 1.0 M aq NH₄OH) gave 25 as a white solid (12 mg, quant, >99:1 dr);^{9b} mp >250 °C; $[\alpha]_{D}^{20}$ -11.5 (c 0.3 in H₂O); {lit.^{9b} $[\alpha]_D^{20} - 14$ (c 0.4 in H₂O)}; ν_{max} (ATR) 3360 (O–H, N–H); $\delta_{\rm H}$ (400 MHz, D₂O) 2.50 (1H, ddd, J 9.7, 5.8, 3.0, C(5)H), 2.84 (1H, dd, J 14.0, 2.4, C(1)H_A), 3.00 (1H, dd, J 14.0, 2.4, $C(1)H_{\rm B}$, 3.29–3.33 (1H, m, C(2)H), 3.44 (1H, t, J 9.7, C(4)H), 3.63-3.68 (2H, m, C(3)H, C(6)H_A), 3.77 (1H, dd, J 11.7, 3.0, $C(6)H_B$; δ_C (100 MHz, D₂O) 46.4 (C(1)), 51.2 (C(2)), 60.9 (C(5)), 60.9 (C(6)), 68.0 (C(4)), 73.1 (C(3)); m/z (ESI⁺) 163 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_6H_{15}N_2O_3^+$ ([M + H]⁺) requires 163.1077; found 163.1078.

(2S,3S,4R,5R)-2-Amino-1,2,5-trideoxy-1,5-imino-D-allose [(+)-2-Amino-1-deoxyallonojirimycin (ADANJ)] 26. Step 1. A solution of 72 (30 mg) in 6.0 M aq HCl (1 mL) and MeOH (1 mL) was stirred at 40 °C for 16 h before being concentrated in vacuo to give (2R,3R,4S,5S)-N(1)-benzyl-2-hydroxymethyl-3,4-dihydroxy-5aminopiperidine hydrochloride as a yellow oil (19 mg); $[\alpha]_D^{20}$ -16.8 (c 0.2 in MeOH); ν_{max} (ATR) 3360 (O-H, N-H), 2942, 2865 (C-H); $\delta_{\rm H}$ (400 MHz, MeOH- d_4) 3.20 (1H, dd, J 11.7, 4.3, C(6) $H_{\rm A}$), 3.25-3.30 (1H, m, C(6)H_B), 3.41-3.46 (1H, m, C(2)H), 3.62-3.66 (1H, m, C(5)H), 3.96 (1H, app d, J 9.8, C(3)H), 4.16-4.18 (1H, m, C(4)H), 4.23-4.26 (2H, m, C(2)CH₂), 4.32 (1H, d, J 11.8, NCH_AH_BPh), 4.86 (1H, d, J 11.8, NCH_AH_BPh), 7.50-7.66 (5H, m, *Ph*); δ_{C} (100 MHz, MeOH- d_{4}) 46.7 (C(6)), 48.0 (C(5)), 54.9 (C(2)CH₂), 58.7 (NCH₂Ph), 63.8 (C(2)), 66.5 (C(3)), 67.7 (C(4)), 130.7, 131.7, 133.0 (o,m,p-Ph), 133.5 (i-Ph); m/z (ESI⁺) 253 ([M + H]⁺, 100%); HRMS (ESI⁺) $C_{13}H_{21}N_2O_3^+$ ([M + H]⁺) requires 253.1547; found 253.1540.

Step 2. $Pd(OH)_2/C$ (10 mg) was added to a stirred solution of the residue of (2R,3R,4S,5S)-N(1)-benzyl-2-hydroxymethyl-3,4-dihydroxy-5-aminopiperidine hydrochloride (19 mg) in degassed MeOH (2 mL), and the resultant suspension was stirred at rt for 48 h under an atmosphere of H₂ (5 atm). HCl (1.0 M in Et₂O, 1 mL) was then added, and the resultant suspension was stirred for a further 5 min before being filtrated through Celite (eluent MeOH) and concentrated in vacuo. Purification via ion exchange chromatography on Dowex-50WX8 resin (hydrogen form, 100-200 mesh, eluent 1.0 M aq NH₄OH) gave 26 as a white solid (12 mg, quant from 72, >99:1 dr); mp > 250 °C; $[\alpha]_{\rm D}^{20}$ +16.3 (c 0.3 in H₂O); $\nu_{\rm max}$ (ATR) 3360 (O–H, N–H); $\delta_{\rm H}$ (400 MHz, D₂O) 2.52 (1H, app t, J 11.8, C(1)H_A), 2.69 (1H, ddd, J 10.4, 5.8, 2.8, C(5)H), 2.76 (1H, dd, J 12.3, 4.8, C(1)H_B), 2.79–2.85 (1H, m, C(2)H), 3.42 (1H, dd, J 10.4, 2.8, C(4)H), 3.59 (1H, dd, J 11.7, 5.8, C(6)H_A), 3.76 (1H, dd, J 11.7, 2.8, C(6)H_B), 3.93-3.95 (1H, m, C(3)H); $\delta_{\rm C}$ (100 MHz, D₂O) 44.6 (C(1)), 50.1 (C(2)), 54.5 (C(5)), 61.5 (C(6)), 69.1 (C(4)), 71.4 (C(3)); m/z (ESI⁺) 163 $([M + H]^+, 100\%);$ HRMS (ESI^+) C₆H₁₅N₂O₃⁺ $([M + H]^+)$ requires 163.1077; found 163.1076.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b01107.

¹H and ¹³C NMR spectra (PDF) Crystallographic information file (for structures CCDC 1479041–1479043) (CIF)

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Notes

The authors declare no competing financial interest.

REFERENCES

(1) Moss, G. P.; Smith, P. A. S. D.; Tavernier, D. Pure Appl. Chem. 1995, 67, 1307.

(2) Compain, P.; Martin, O. R. Iminosugars: past, present and future. In *Iminosugars: From Synthesis to Therapeutic Applications;* Compain, P., Martin, O. R., Eds.; Wiley: Chichester, 2007; pp 1–6.

(3) Anderson, J. W.; Nicolosi, R. J.; Borzelleca, J. F. Food Chem. Toxicol. 2005, 43, 187.

(4) Iida, H.; Yamazaki, N.; Kibayashi, C. J. Org. Chem. 1987, 52, 3337.

(5) Hlavka, J. J.; Child, R. G.; Bitha, P.; Lin, Y. I. U.S. Patent 4587331, 1986.

(6) Wrodnigg, T. M.; Diness, F.; Gruber, C.; Hausler, H.; Lundt, I.; Rupitz, K.; Steiner, A. J.; Stütz, A. E.; Tarling, C. A.; Withers, S. G.; Wolfler, H. *Bioorg. Med. Chem.* **2004**, *12*, 3485.

(7) Kappes, E.; Legler, G. J. Carbohydr. Chem. 1989, 8, 371.

(8) Gradnig, G.; Legler, G.; Stutz, A. E. Carbohydr. Res. 1996, 287, 49.

(9) For instance, see: (a) Tsou, E.-L.; Yeh, Y.-T.; Liang, P.-H.; Cheng, W.-C. *Tetrahedron* **2009**, *65*, 93. (b) Le Merrer, Y.; Poitout, L.; Depezay, J.-C.; Dosbaa, I.; Geoffroy, S.; Foglietti, M.-J. *Bioorg. Med. Chem.* **1997**, *5*, 519.

(10) For example, see: (a) Davies, S. G.; Lee, J. A.; Roberts, P. M.; Thomson, J. E.; West, C. J. *Tetrahedron* **2012**, *68*, 4302. (b) Davies, S. G.; Foster, E. M.; Lee, J. A.; Roberts, P. M.; Thomson, J. E. *Tetrahedron* **2013**, *69*, 8680.

(11) For example, see: (a) Bagal, S. K.; Davies, S. G.; Lee, J. A.; Roberts, P. M.; Scott, P. M.; Thomson, J. E. *J. Org. Chem.* **2010**, *75*, 8133. (b) Davies, S. G.; Fletcher, A. M.; Lee, J. A.; Roberts, P. M.; Russell, A. J.; Taylor, R. J.; Thomson, A. D.; Thomson, J. E. *Org. Lett.* **2012**, *14*, 1672.

(12) For example, see: (a) Brock, E. A.; Davies, S. G.; Lee, J. A.; Roberts, P. M.; Thomson, J. E. Org. Lett. **2012**, *14*, 4278. (b) Davies, S. G.; Fletcher, A. M.; Lee, J. A.; Lorkin, T. J. A.; Roberts, P. M.; Thomson, J. E. Org. Lett. **2013**, *15*, 2050.

(13) Davies, S. G.; Fletcher, A. M.; Roberts, P. M.; Thomson, J. E. *Tetrahedron: Asymmetry* **2012**, 23, 1111.

(14) Davies, S. G.; Figuccia, A. L. A.; Fletcher, A. M.; Roberts, P. M.; Thomson, J. E. *Tetrahedron* **2014**, *70*, 3601.

(15) Davies, S. G.; Figuccia, A. L. A.; Fletcher, A. M.; Roberts, P. M.; Thomson, J. E. Org. Lett. **2013**, 15, 2042.

(16) Cook, P. D. U.S. Patent 9810286, 1998.

(17) Solé, D.; Garcia-Rubio, S.; Bosch, J.; Bonjoch, J. Heterocycles 1996, 43, 2415.

(18) The assigned identity of **32** was supported by ¹H and ¹³C NMR spectroscopic analyses; furthermore, the IR spectrum of **32** displayed a diagnostic C==O absorbance at 1799 cm⁻¹ that confirmed the presence of a carbonate moiety.

(19) The ¹H NMR spectrum of **33** was also recorded in CH_2Cl_2 at low temperature (208 K); however, even though two conformers of **33** were observed in this spectrum, the ¹H NMR ³*J* coupling constants of these two conformers could not be resolved due to insufficient dispersion of peaks.

(20) Only one reference containing the ¹H NMR characterization data for **33** has been reported in the literature, see: Ortiz, A.; Young, I. S.; Sawyer, J. R.; Hsiao, Y.; Singh, A.; Sugiyama, M.; Corbett, R. M.;

Chau, M.; Shi, Z.; Conlon, D. A. Org. Biomol. Chem. 2012, 10, 5253. (21) Only one reference containing the ¹H NMR characterization data for 36 has been reported in the literature, see: Terentiev, P. B.; Zilberstein, T. M.; Borisenko, A. A.; Shmorgunov, V. A.; Piskunkova, N. F.; Grishina, G. V. Chem. Heterocycl. Compd. 2003, 39, 885.

(22) Pavlova, M.; Hoenke, C.; Christoffers, J. Synthesis 2009, 1659. (23) Csatayová, K.; Davies, S. G.; Ford, J. G.; Lee, J. A.; Roberts, P.

(25) Csatayova, K.; Davies, S. G.; Fold, J. G.; Lee, J. A.; J. M.; Thomson, J. E. J. Org. Chem. **2013**, 78, 12397.

(24) VanRheenen, V.; Kelly, R. C.; Cha, D. Y. Tetrahedron Lett. 1976, 17, 1973.

(25) Crystallographic data (excluding structure factors) for the structures of **38**, **56**, and **69** have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication numbers CCDC 1479041, 1479042, and 1479043, respectively.

(26) The ¹³C NMR chemical shift for CH₂I carbons in pyrrolidines are typically lower ($\delta_{\rm C} = 1.5-12$ ppm) than the ¹³C NMR chemical shift for CHI carbons in piperidines ($\delta_{\rm C} = 20-40$ ppm), for example, see: Davies, S. G.; Nicholson, R. L.; Price, P. D.; Roberts, P. M.; Russell, A. J.; Savory, E. D.; Smith, A. D.; Thomson, J. E. *Tetrahedron: Asymmetry* **2009**, *20*, 758.

(27) The C(4)*H* proton was not observed in the ¹H NMR spectrum of **39** in CDCl₃ at rt.

(28) The atomic connectivities within 55 and 56 were initially assigned by ¹H and ¹³C NMR spectroscopic analyses, and the IR spectra of 55 and 56 also displayed diagnostic absorbances at ~1780 cm⁻¹ for the C=O of the carbamate moiety.

(29) The assigned bicyclic system within 57 was entirely consistent with both 1 H and 13 C NMR spectroscopic analyses of 57, and the IR

spectrum of 57 displayed a diagnostic C=O absorbance at 1797 cm⁻¹ for the carbamate moiety.

(30) An impurity that was tentatively assigned as N(1)- α -methylbenzyl-substituted pyrrolidine **62** was observed in the ¹H NMR spectrum of this sample of **12** [$\delta_{\rm H}$ = 1.5 ppm (3H, d, C(α)Me) and $\delta_{\rm H}$ = 4.12 ppm (1H, q, C(α)H)].

(31) (a) Flack, H. D. Acta Crystallogr., Sect. A: Found. Crystallogr. 1983, 39, 876. (b) Flack, H. D.; Bernardinelli, G. J. Appl. Crystallogr. 2000, 33, 1143.

(32) Llaveria, J.; Beltrán, A.; Sameera, W. M. C.; Locati, A.; Díaz-Requejo, M. M.; Matheu, M. I.; Castillón, S.; Maseras, F.; Pérez, P. J. J. Am. Chem. Soc. **2014**, 136, 5342.

(33) Pangborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. J. Organometallics **1996**, *15*, 1518.

(34) Betteridge, P. W.; Carruthers, J. R.; Cooper, R. I.; Prout, C. K.; Watkin, D. J. J. Appl. Crystallogr. **2003**, *36*, 1487.

(35) Khamrai, U.; Karak, S. K.; Ronsheim, M.; Saha, A. K. U.S. Patent 0168080 A1 20100701, 2010.

(36) Grishina, G. V.; Borisenko, A. A.; Veselov, I. S.; Petrenko, A. M. Russ. J. Org. Chem. **2005**, 41, 272.

(37) Compound 37 was found to decompose upon attempted purification via flash column chromatography. The crude reaction mixture was therefore characterized as a 50:50 mixture of 37 and DMAP.

(38) A peak corresponding to C(4)H was not observed in the ¹H NMR spectrum of **38** in CDCl₃ at rt.

(39) The peaks in the ¹³C NMR spectrum of 38 corresponding to C(3) and C(6) at 39.9 and 48.3 ppm, respectively, were extremely broad.

(40) Rives, A.; Génisson, Y.; Faugeroux, V.; Zedde, C.; Lepetit, C.; Chauvin, R.; Saffon, N.; Andrieu-Abadie, N.; Colié, S.; Levade, T.; Baltas, M. *Eur. J. Org. Chem.* **2009**, 2474.

(41) Roush, W. R.; Gustin, D. Tetrahedron Lett. 1994, 35, 4931.